

# Continuous Manufacturing Platform for Lipid and Polymer-based Nanoparticle Therapeutics

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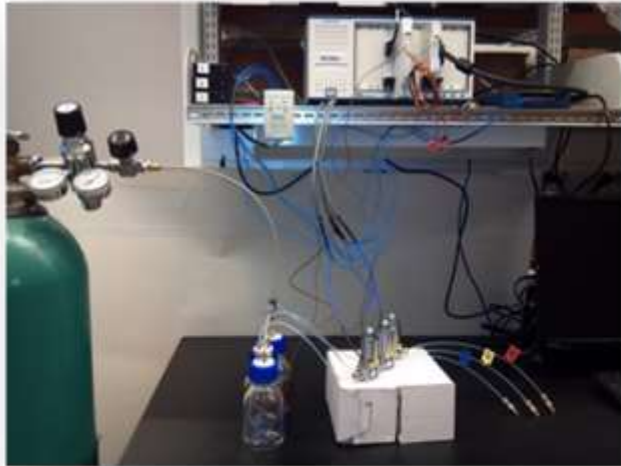
10/11/2023

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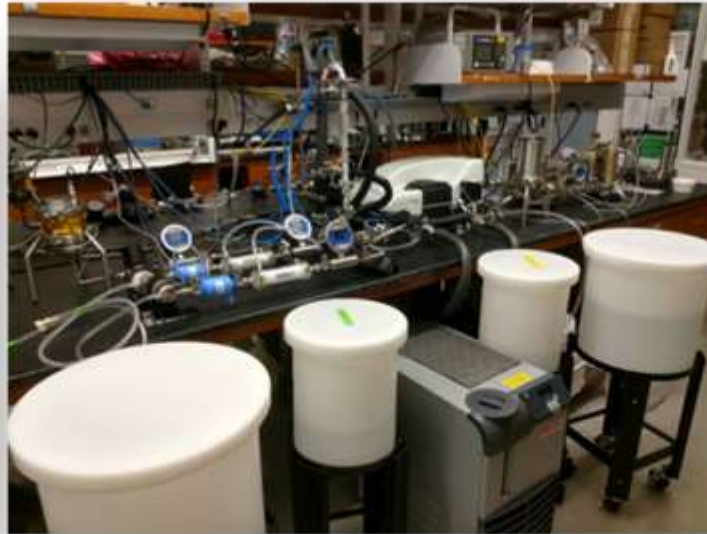
Slide 1

# From laboratory to industrial technology

2014



2018



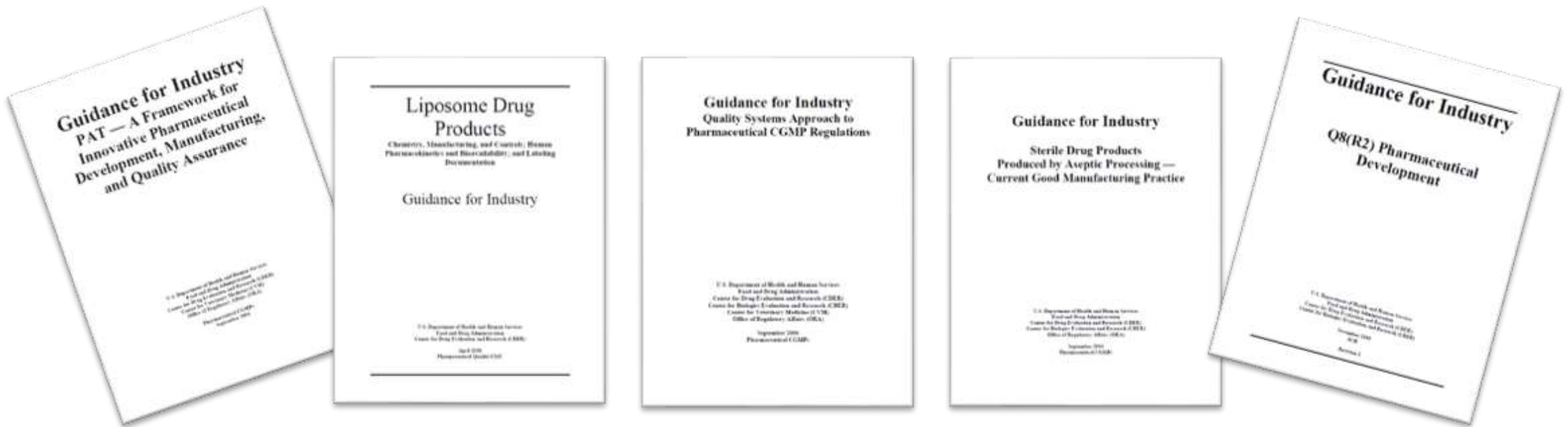
2022



# Modernizing Pharma Manufacturing

“One of today’s most important tools for modernizing the pharmaceutical industry is a process known as continuous manufacturing...”

~Joint Statement by former FDA Commissioner Scott Gottlieb and FDA Director Janet Woodcock, 2019

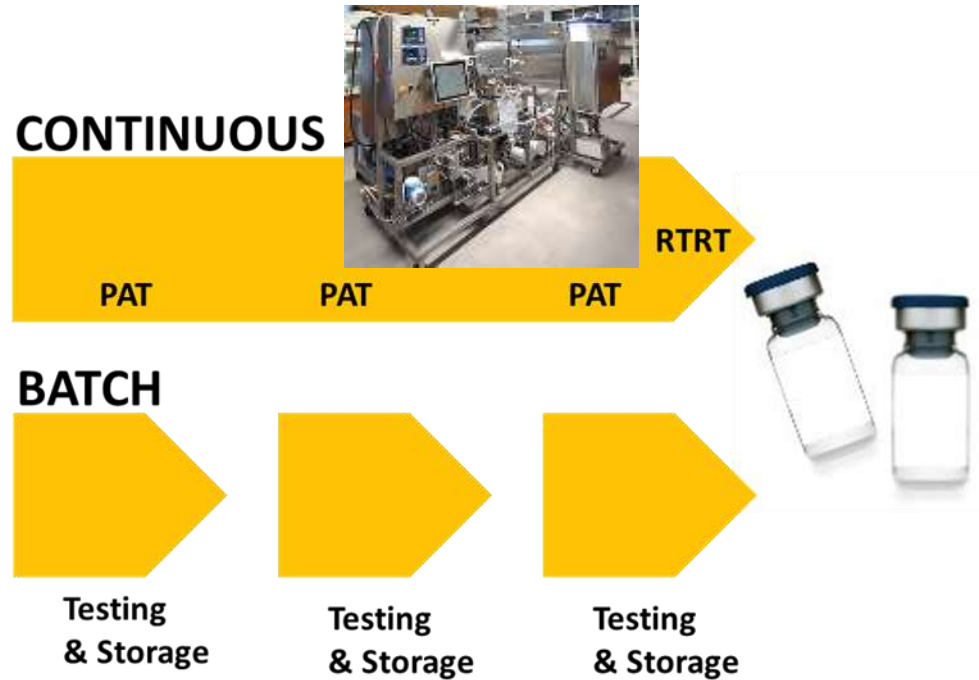


“Allows FDA to issue grants to study continuous manufacturing — a technologically advanced and automated manufacturing method.”

~The 21<sup>st</sup> Century Cures Act, 2019



# Batch vs. Continuous



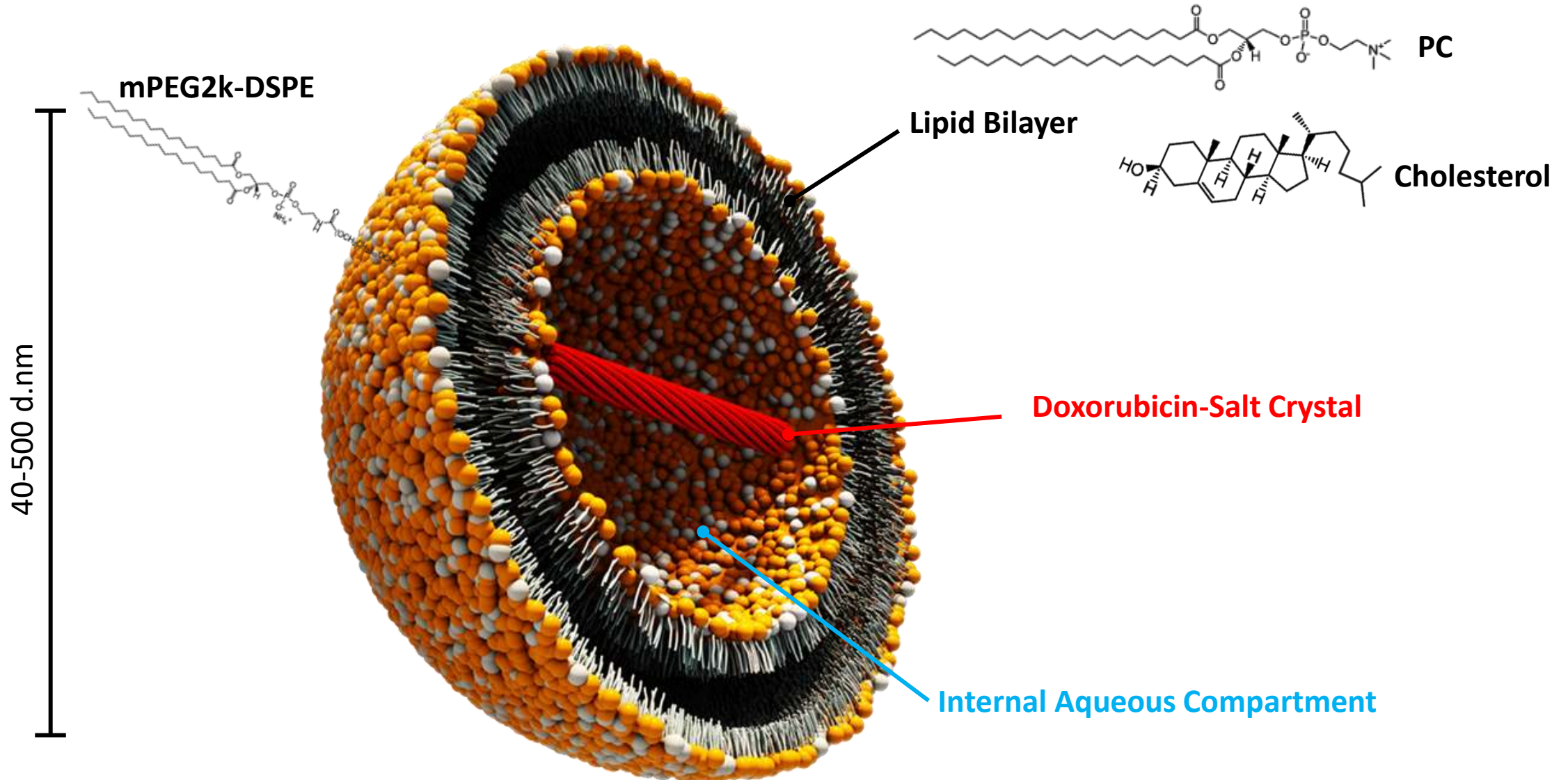
*"...quality cannot be tested into products; it should be built-in or should **be** by design"*

FDA Guidance on PAT Framework

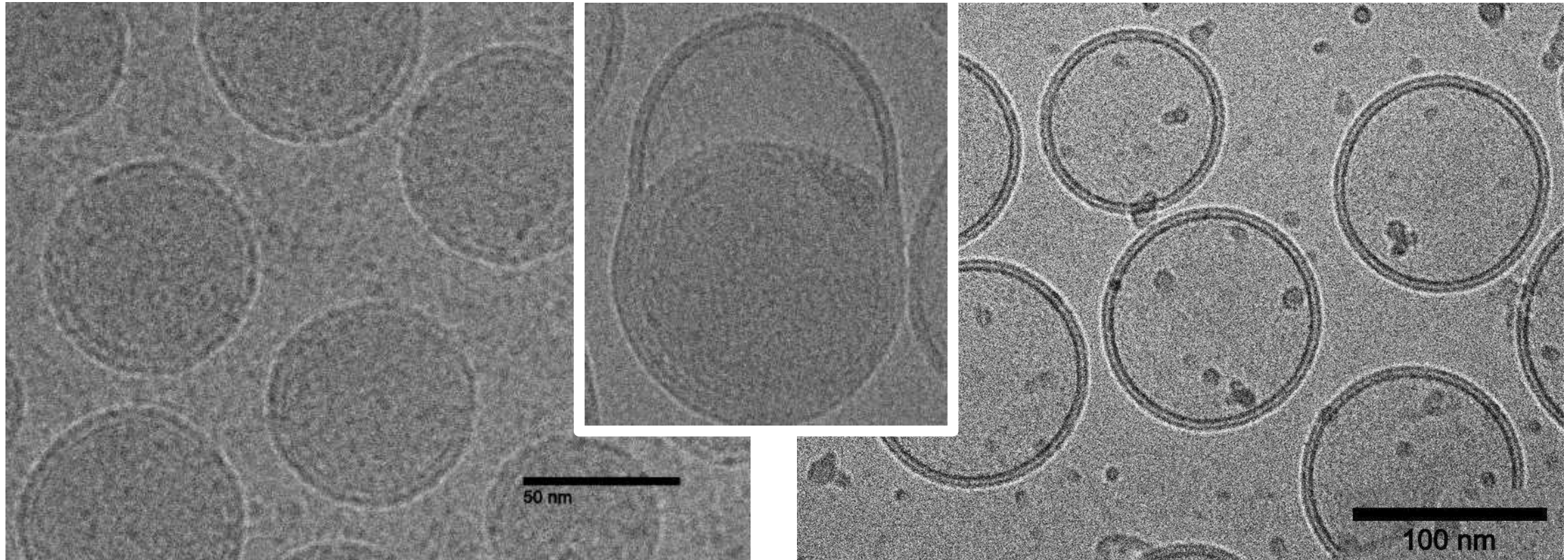
## Continuous Manufacturing Benefits

- **Higher Quality Products**
- **Faster to Market**
- **Scalable**
- **Process Analytical Technology**
  - Fine-Control of Quality Attributes
- **Reduce Product Waste**
  - Divert Process if Error Occurs
- **Reduce System Footprint**
  - Portable System
- **Reduce Human Error**
  - Reduction in Open Transfers
- **Supports End-to-End Manufacturing**

# Liposomal Nanoparticle Properties and Structure



# LNP vs Liposome Structure



## *Lipid Nanoparticles*

1. PC Lipid .....
2. Cholesterol .....
3. Pegylated Lipid .....
4. Ionizable Lipid
5. Nucleic Acid

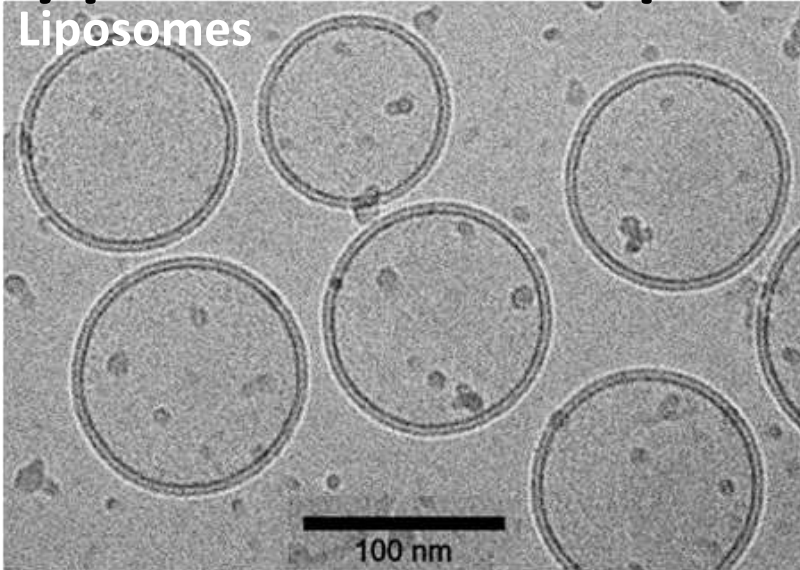
## *Liposomes*

1. PC Lipid
2. Cholesterol
3. Pegylated Lipid

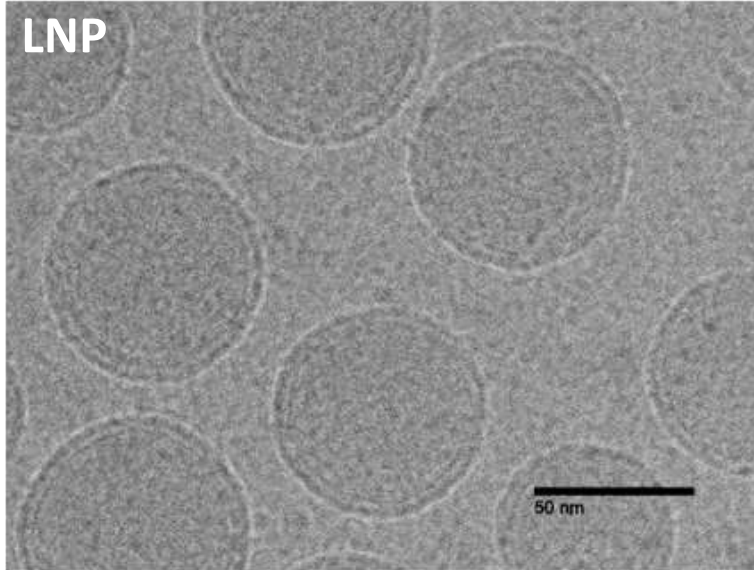


# Types of Nanoparticles

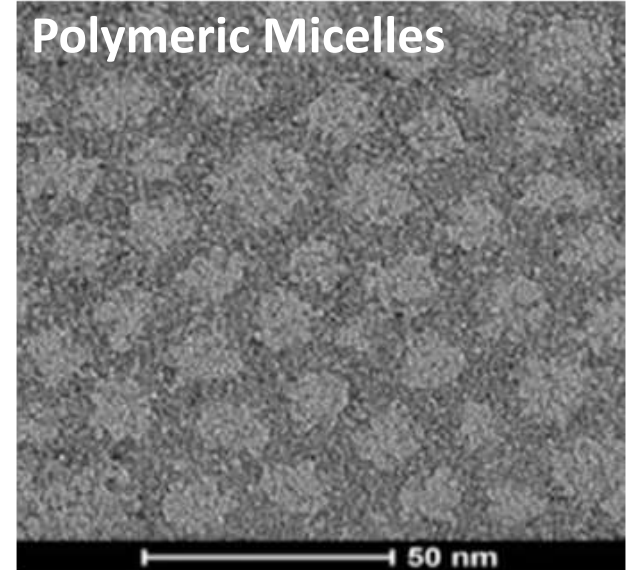
Liposomes



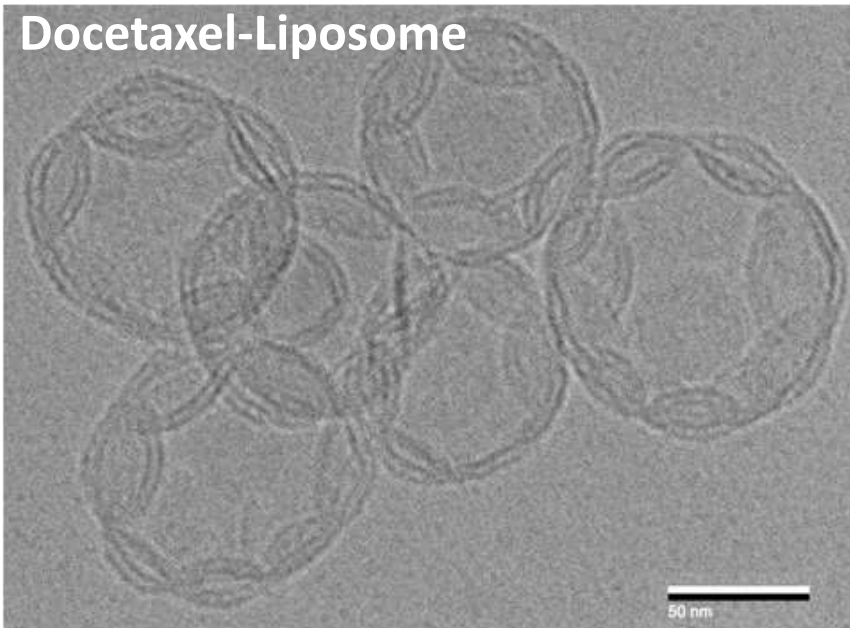
LNP



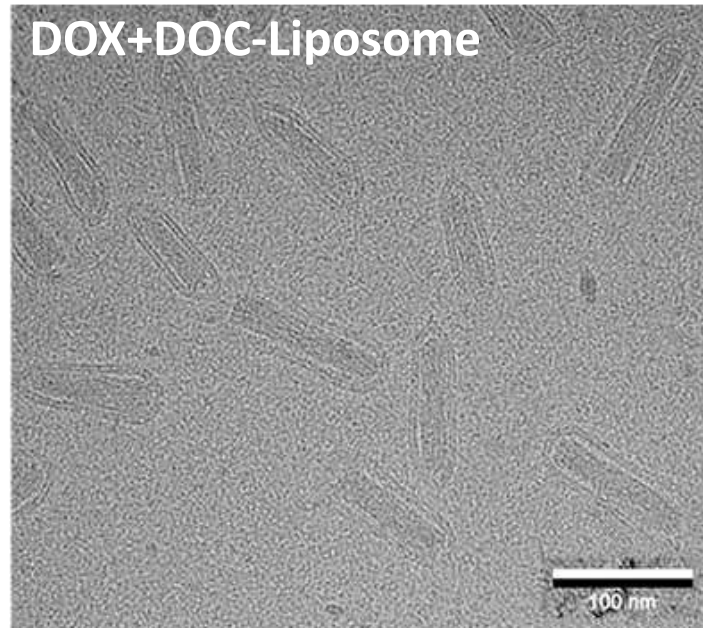
Polymeric Micelles



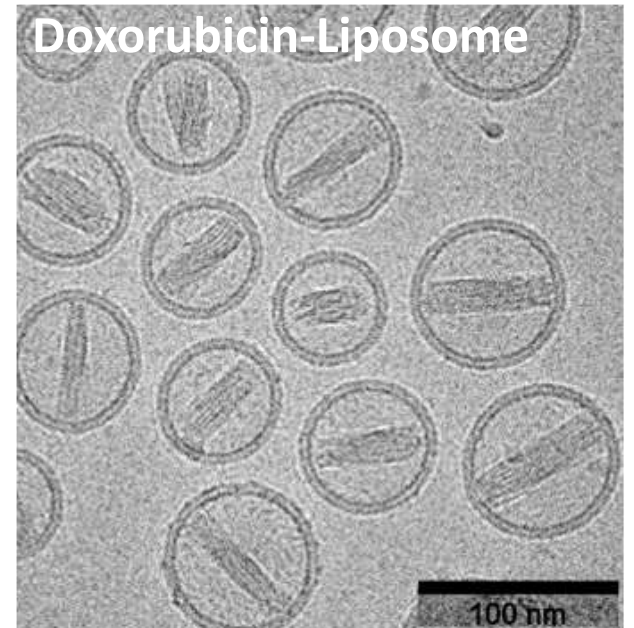
Docetaxel-Liposome



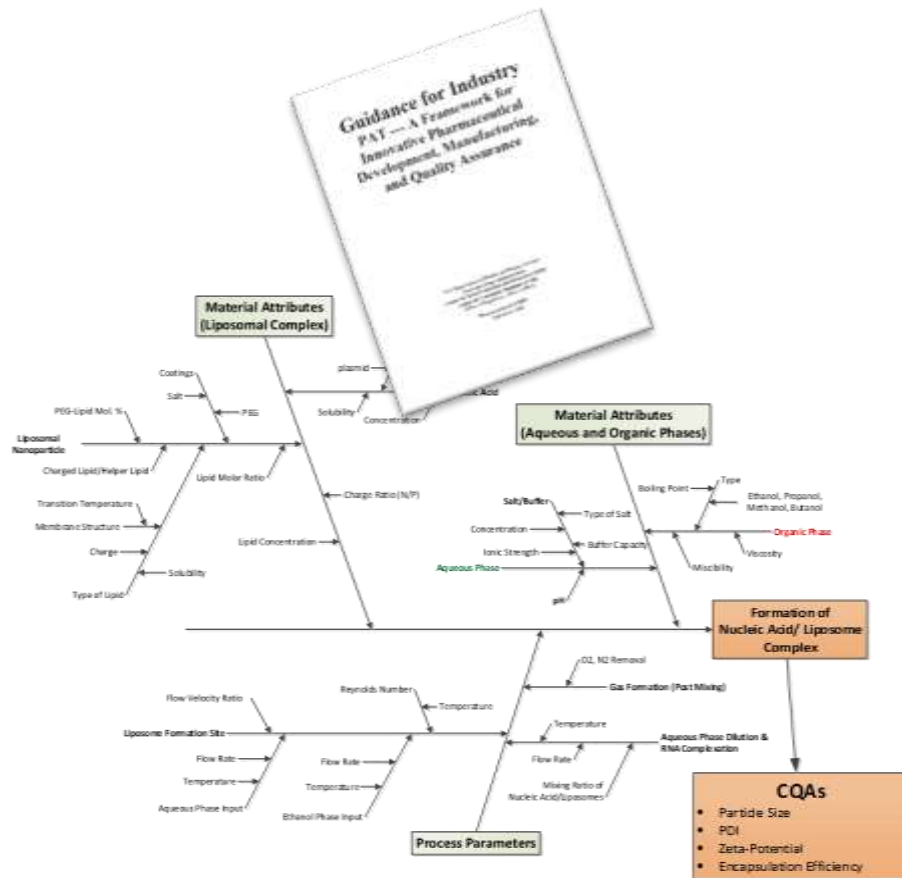
DOX+DOC-Liposome



Doxorubicin-Liposome



# Quality by Design - A risk-based approach to the problem



*Example Ishikawa Diagram to relate material attributes and process parameters to cQAs.*

## Material Attributes

- Lipid Concentration
- Lipid Molar Ratios
- Lipid Purity
- Lipid pKa
- Lipid Headgroup
- Lipid Ionizable Species
- Aqueous Phase
  - Salt Additions
  - Organic Phase Additions

## Process Parameters

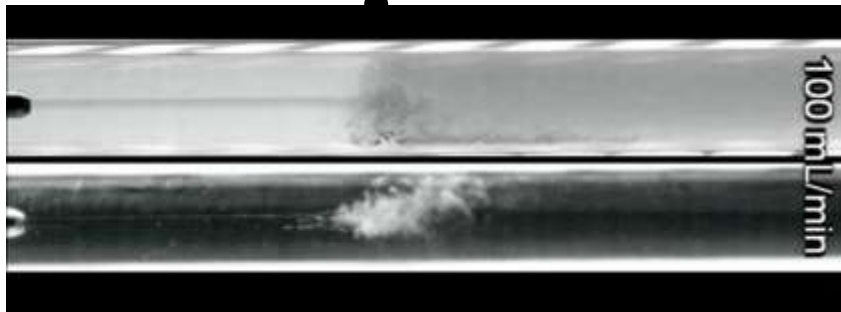
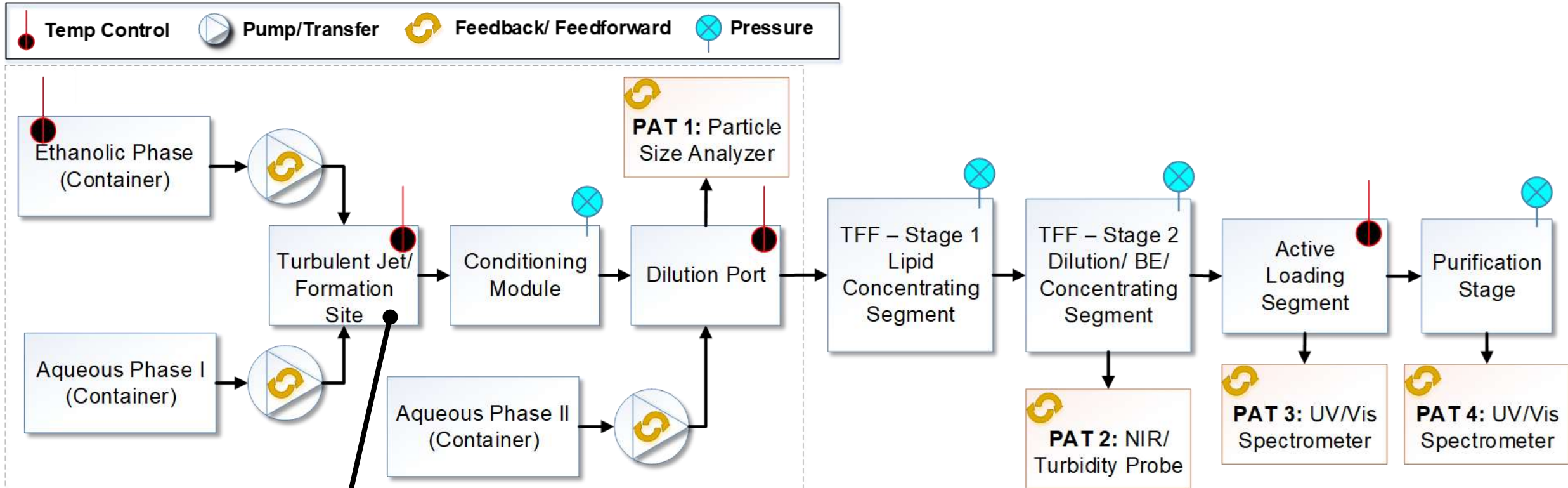
- Aqueous Flow Rates
- Ethanol Flow Rates
- Reynolds Number of Mixing
- Aqueous Phase Temperature
- Ethanolic Phase Temperature
- pH of Aqueous Phase

### Quality Attributes

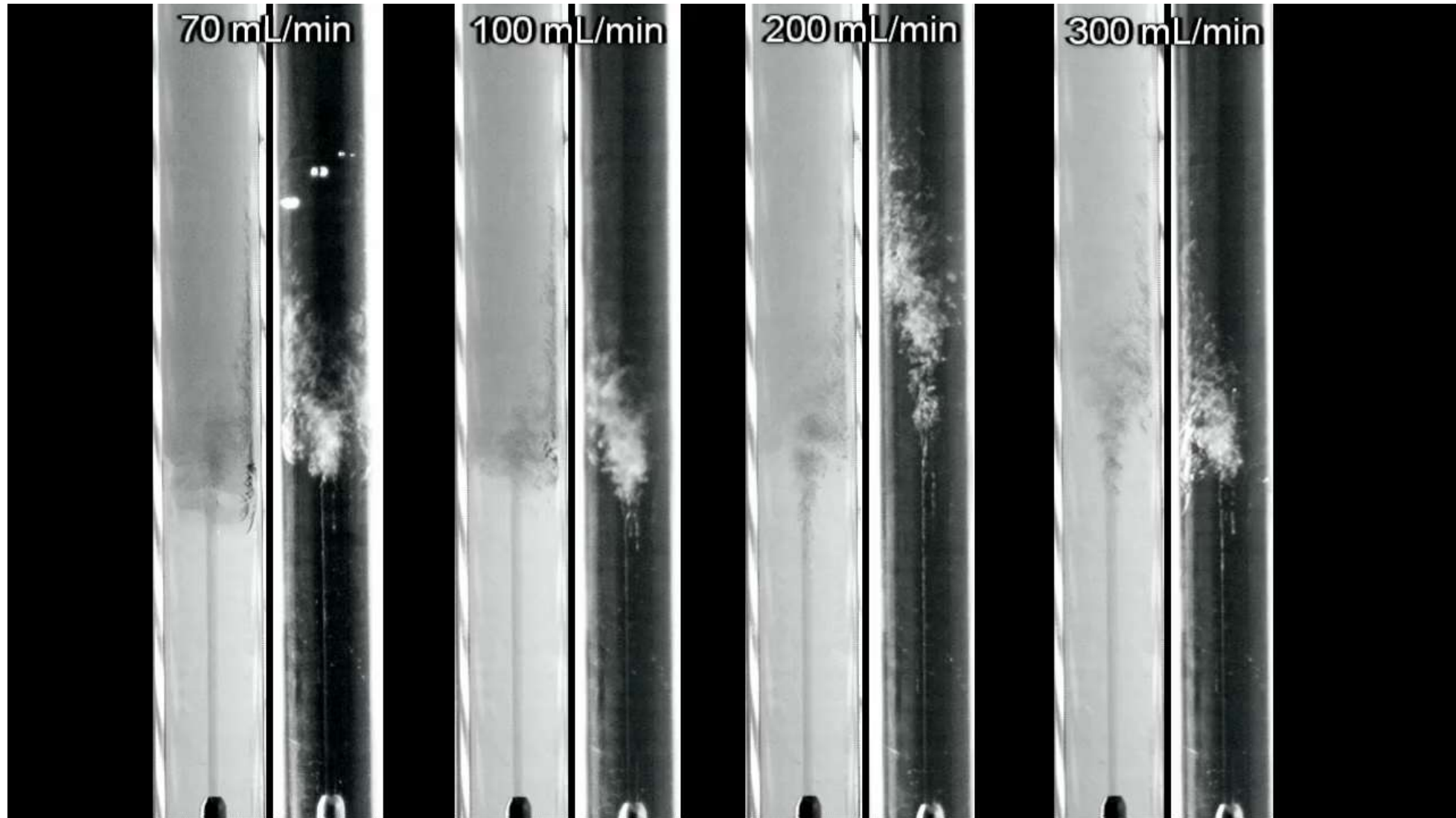
- Particle Size
- Polydispersity Index (PDI)
- Zeta-Potential and Deviation
- Encapsulation Efficiency
- Total Drug
- Drug Loading
- Drug Crystal Structure
- Residual Solvent



# Nanoparticle Process Flow Chart

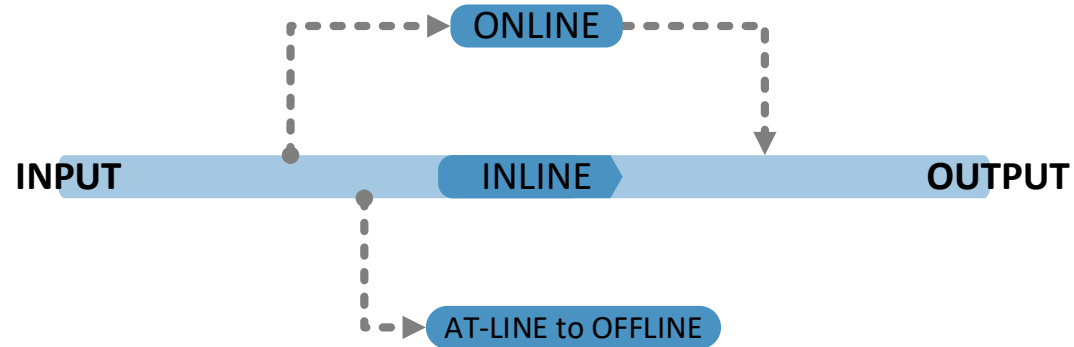


# High-Speed Camera: Jet Formation



Entire 30 second video takes place in less than 1 second...

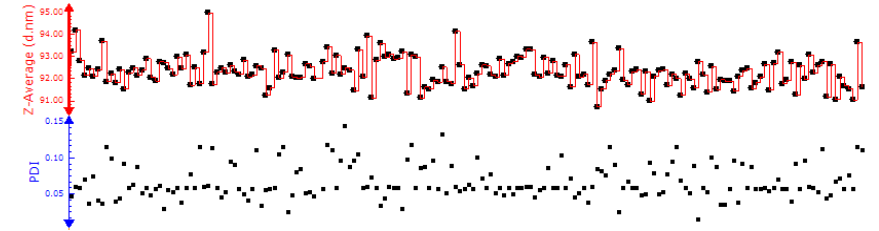
# Process Analytical Technology



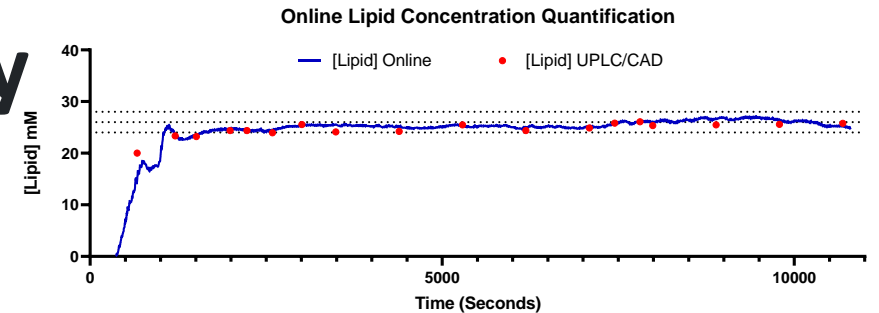
**PAT Tools:** Process analyzer or one or more “soft sensors” with predictive algorithms (multivariate) to determine critical attributes.

**DLS**

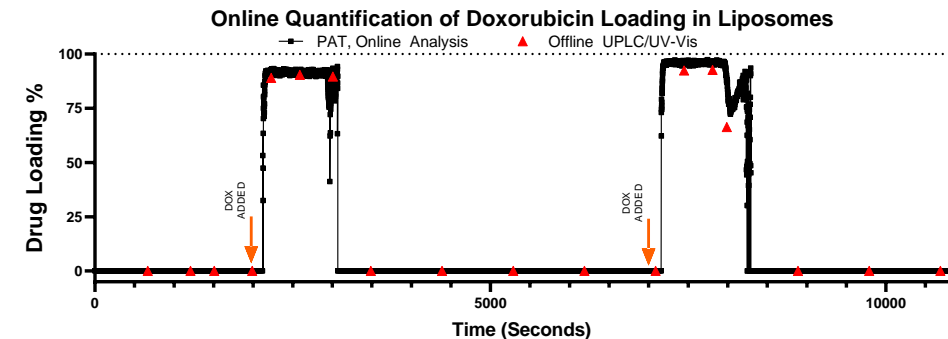
Z-Average  
PDI



**Turbidity**  
[Particle]

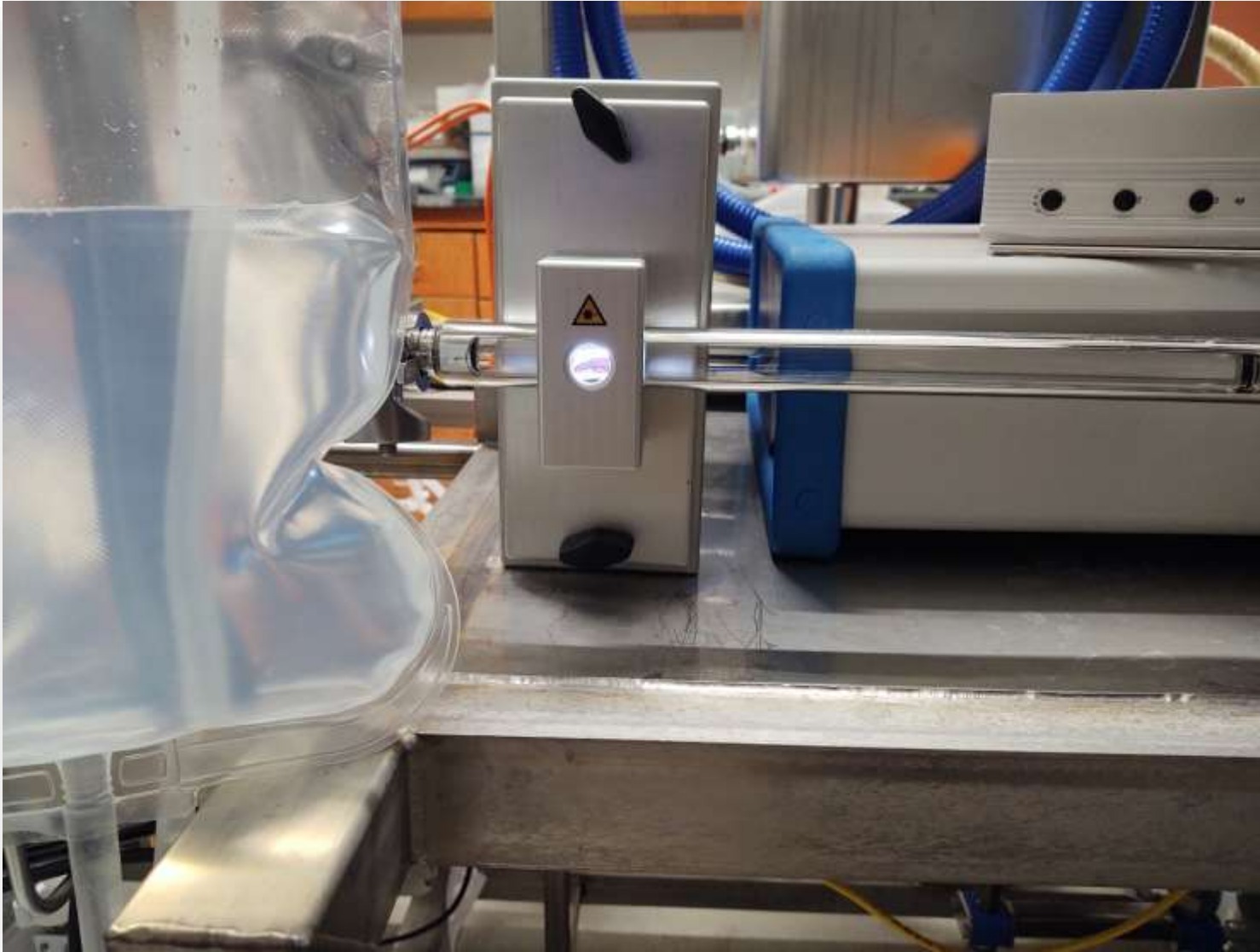


**UV/Vis**  
[Drug]  
%E





# PAT Integration: Particle Size Setup



## NanoFlowSizer

Spatially Resolved-DLS

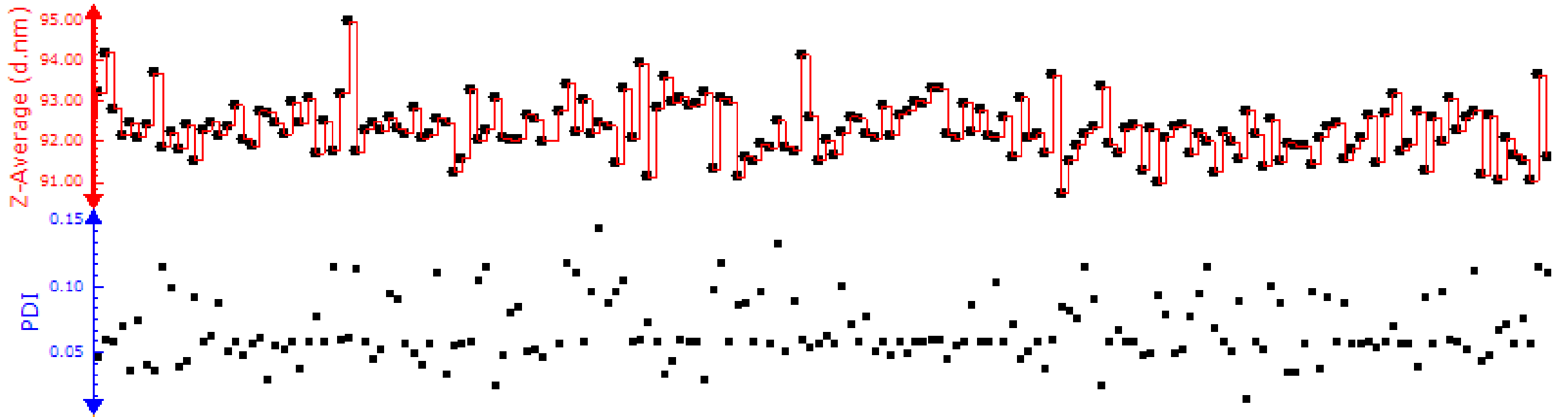
Mode: Online

Measurement Interval: <6s

### Benefits

- Rapid, online analysis
- Measurements at high flow rates
- Low and high concentrations acceptable
- Compliant Software package

# Monitor and Control of Particle Size



## Measured Attributes

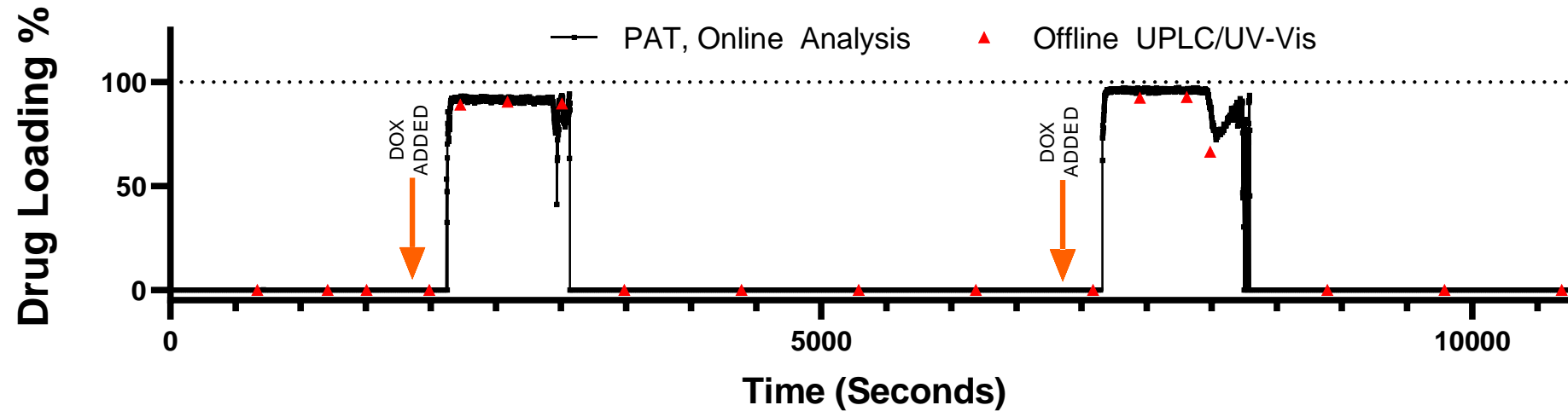
1. Z-Average (d.nm)
2. CumulantPDI
3. Shear Rate
4. Temperature

Relate intermediate liposomes at formation site to liposomes as the “end-product”.

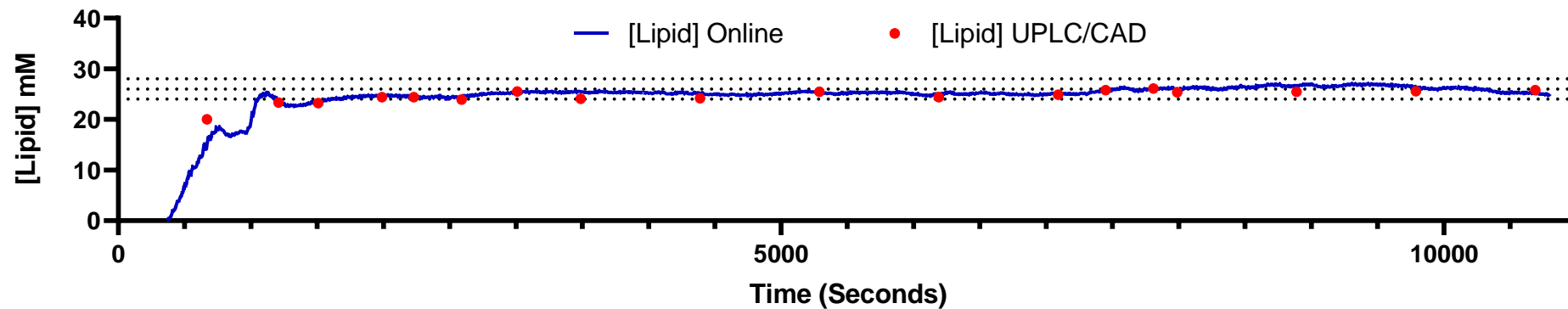
**Predictive behavior of intermediates to “end-product”?**

# Online Doxorubicin and Lipid Concentration Analysis

## Online Quantification of Doxorubicin Loading in Liposomes



## Online Lipid Concentration Quantification





# DOE Example: Material Attributes

**Multi-factorial Design (5x3x2x3x3 = 270 runs)**

**Lipid Formulation:** (Main Lipid Type):Chol:DSPG

DSPG was fixed at 5% molar ratio.

## Factors

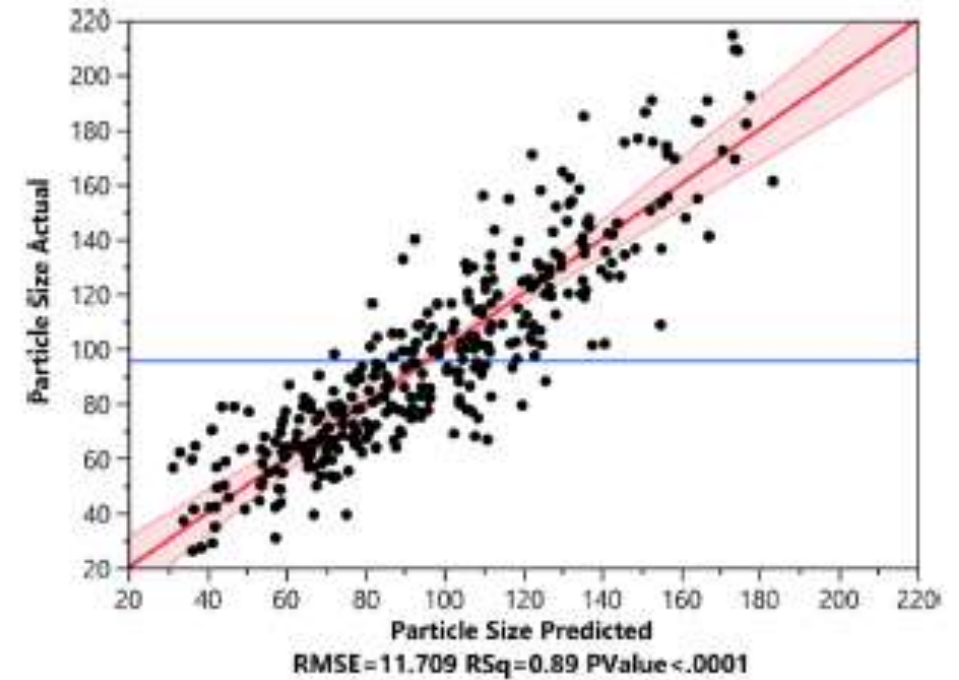
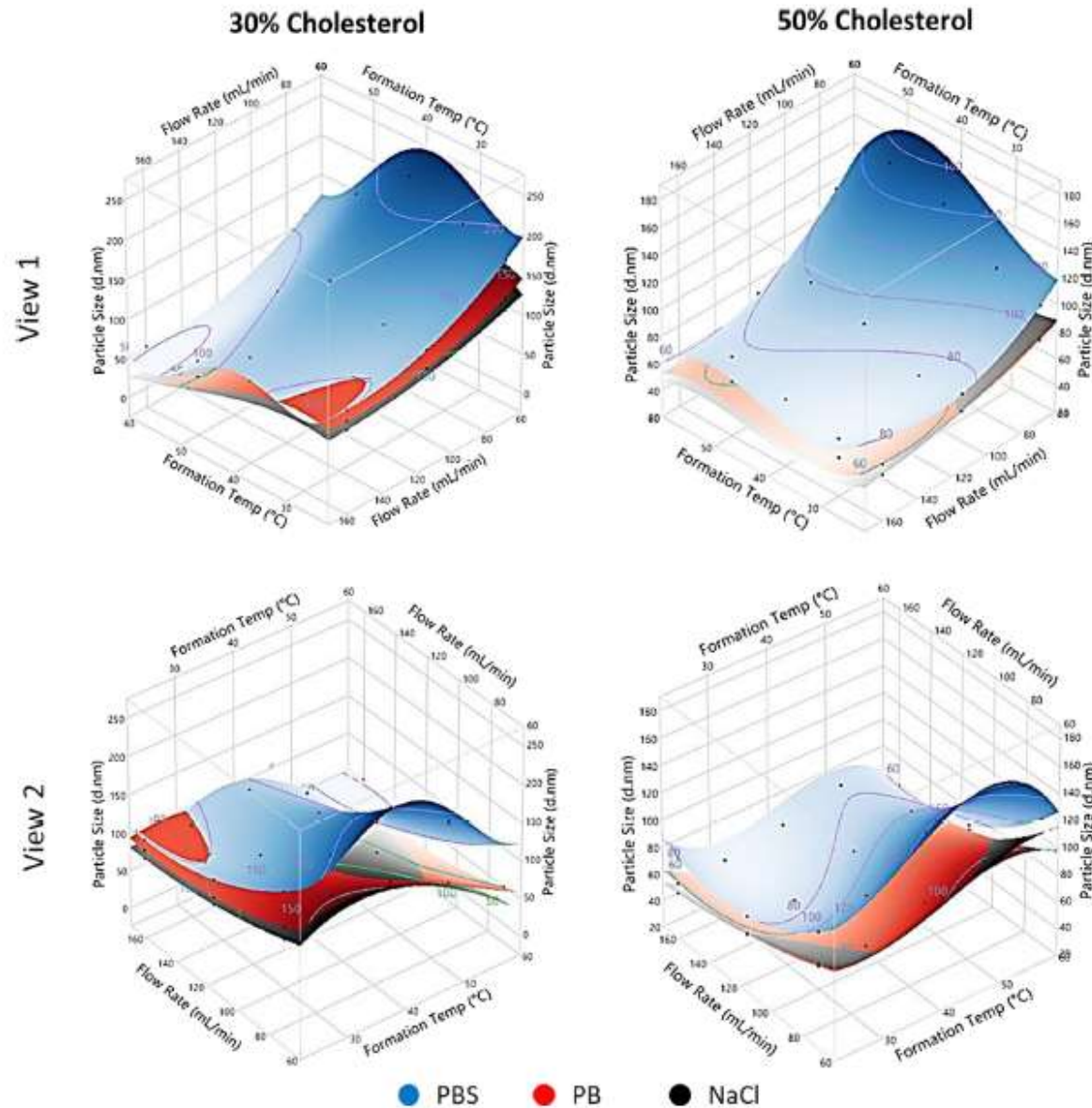
- Post Formation Temperature (20-60°C)
- Aqueous Phase Flow Rate (70-160 mL/min)
- Cholesterol Percentage (30-50%)
- Main Lipid Type (**DMPC** vs **DPPC** vs **DSPC**)
- Aqueous Phase Salt (**PBS**, **PB** or **NaCl**)
  - pH set to 7.45 for PBS and PB.



## Responses

- Z-Average Particle Size
- PDI (Polydispersity Index)
- Zeta-Potential

# HSPC-based liposomes



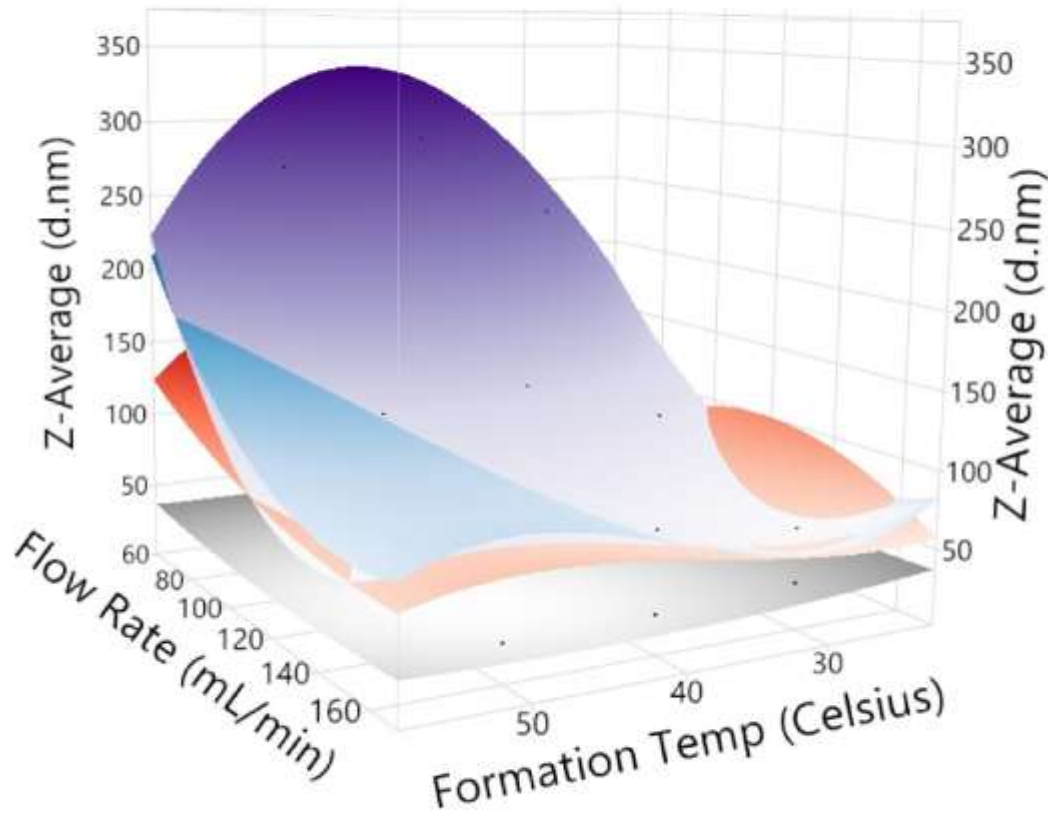
# Type of Lipid on Size

● DOPC

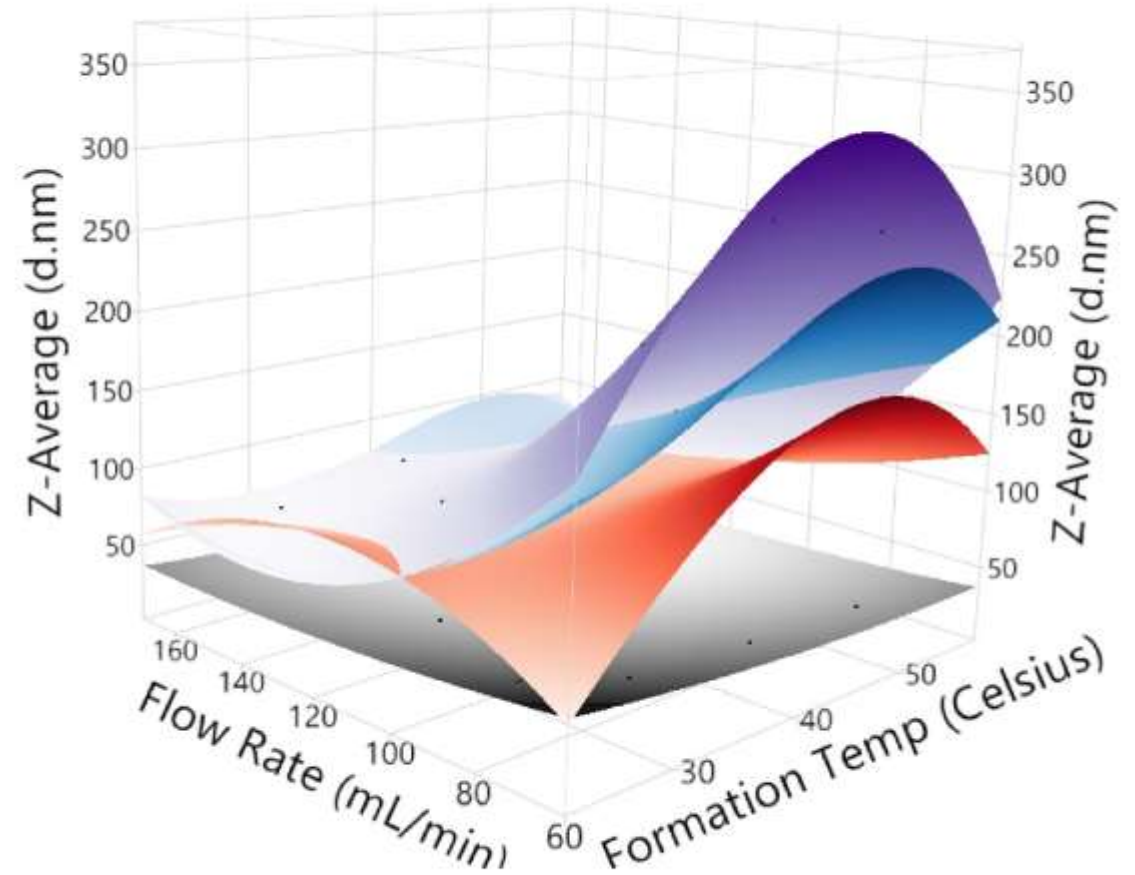
● DMPC

● DPPC

● DSPC



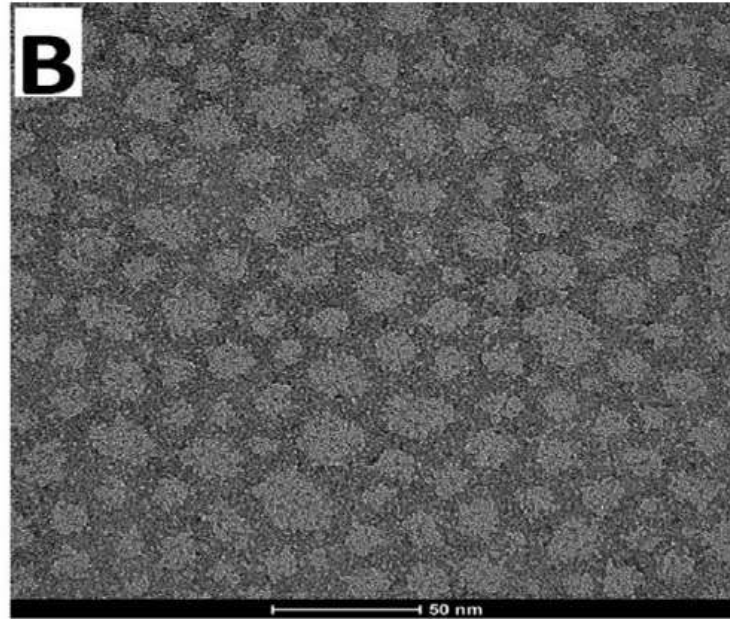
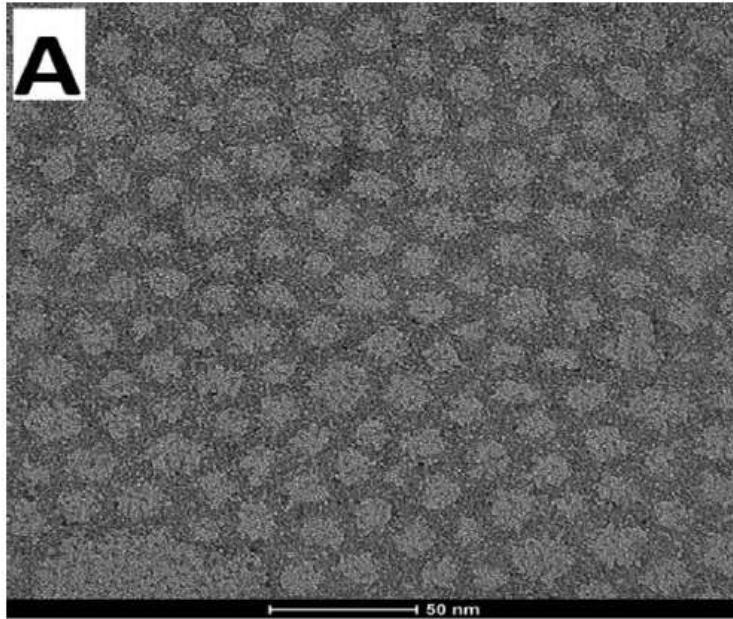
View 1



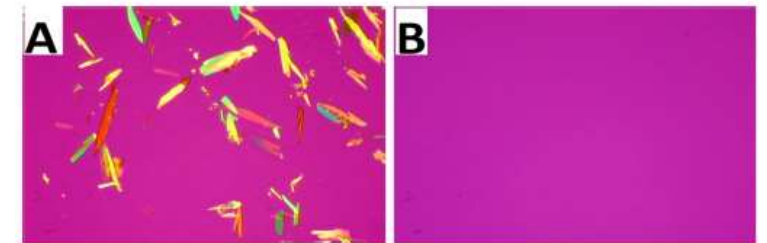
View 2



# Formulation and Characterization of Polymeric Micelles



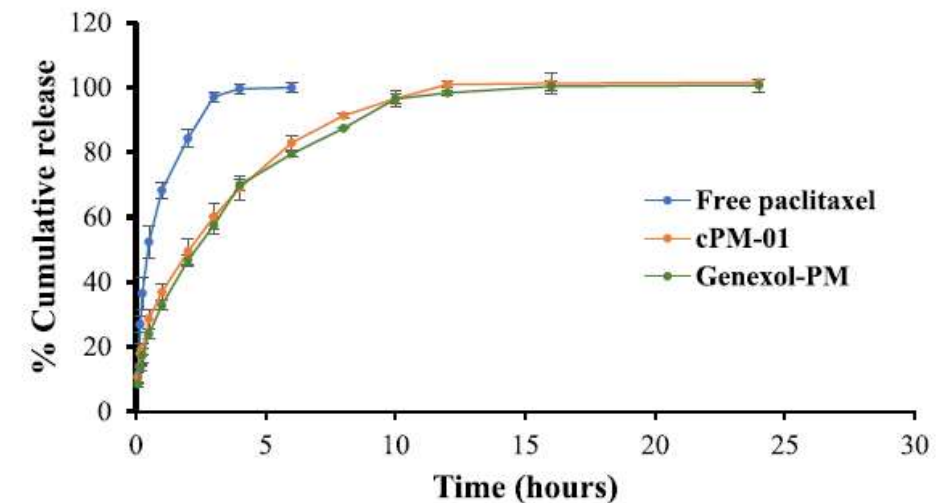
Transmission electron microscopy images of: (A) blank; and (B) curcumin-loaded micelles.



Polarized light microscopy images of: (A) free curcumin (5% ethanol in water); and (B) curcumin-loaded polymeric micelles (5% ethanol in water).



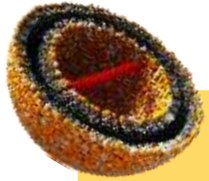
Appearance of: (1) blank polymeric micelles, transparent; (2) curcumin-loaded polymeric micelles, yellow transparent; and (3) free curcumin in water, yellow opaque.



# Case Study: Liposomal Doxorubicin

*Performed at*  
**URI Pharmaceutical Development  
Institute Training Center**

# Liposomal Doxorubicin Specification



Parameters	Brand Name/ Batch Process	Continuous Processing
Lipid composition	HSPC/Chol/mPEG2000-DSPE (56.3:38.4:5.3 mol%)	
[Lipid]	20 mM	
[Doxorubicin-HCl]	2 mg/mL	
Drug Encapsulation	>90% Leaflet 99% Tested	>95% No Purification >99% Post Purification
Loading Battery	Ammonium sulfate 250mM	
External Buffer	10mM histidine, pH 6.5 + 10% Sucrose	
Particle Size (d.nm)	80-85 nm (DLS)	



# System in Clean-Room Suite

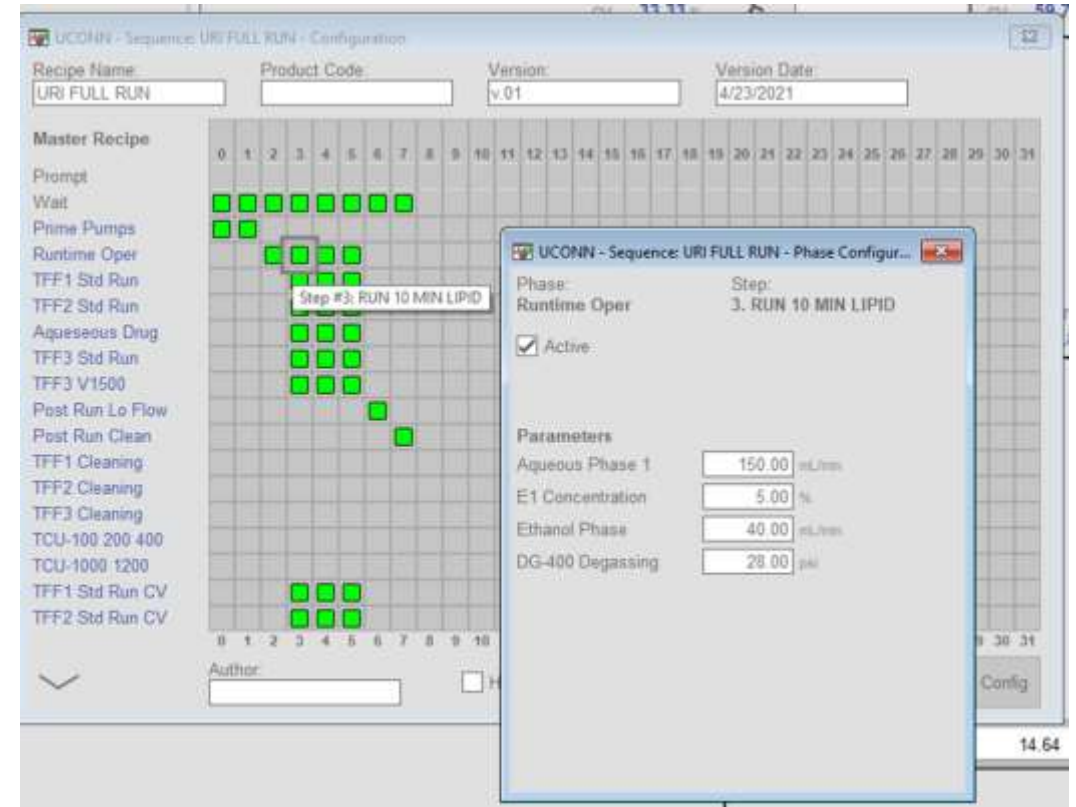


Cleanroom at URI, UConn System front and back views

# Procedures and Overview

1. Gowning procedures
2. Draft batch record
3. Draft cleaning SOP
4. System SOPS
  - Run-time Recipes
5. Environmental monitoring report
6. Cleaning cycle development report
  - Executed UCONN cleaning cycle form
7. Personnel monitoring

Example Run-time Recipe



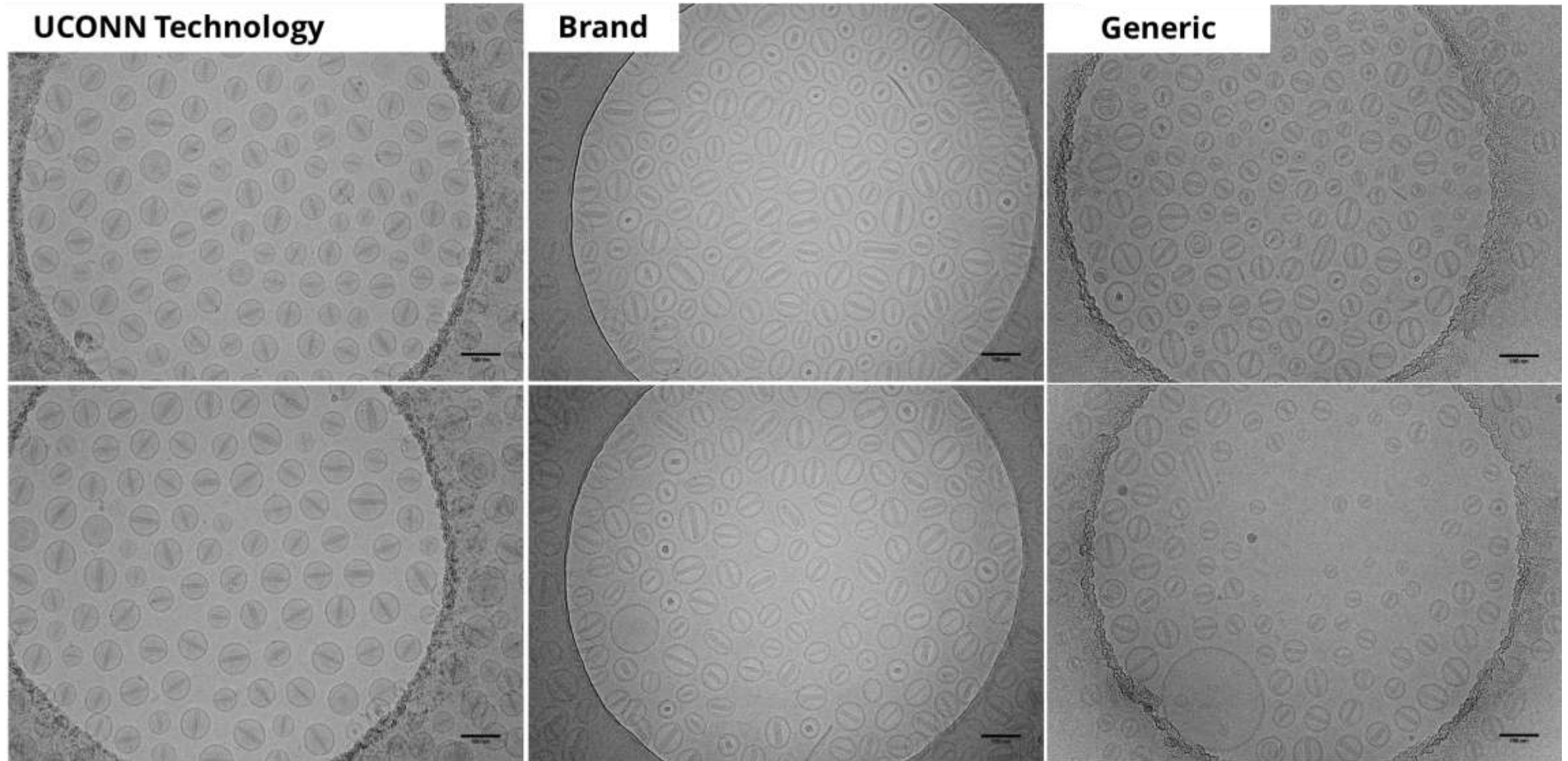
# Final Particle Size Results (n=3)

Sample Name	Z-Ave (d.nm)	PDI	EE (%)
D29t-Run 1	87.54	0.027	85%
D29t-Run 2	86.45	0.035	83%
E5t-Run 3	87.94	0.035	90%
AVERAGES:	87.3	0.032	86.0%
STDEV:	0.771	0.005	3.6%

- Three runs with cleaning in-between each run cycle.
- EE% is the drug encapsulation of doxorubicin in the liposomes.
  - Lower than expected, heating stage was lower.
- Runs were successful.
  - Low standard deviations and percent errors are reported.

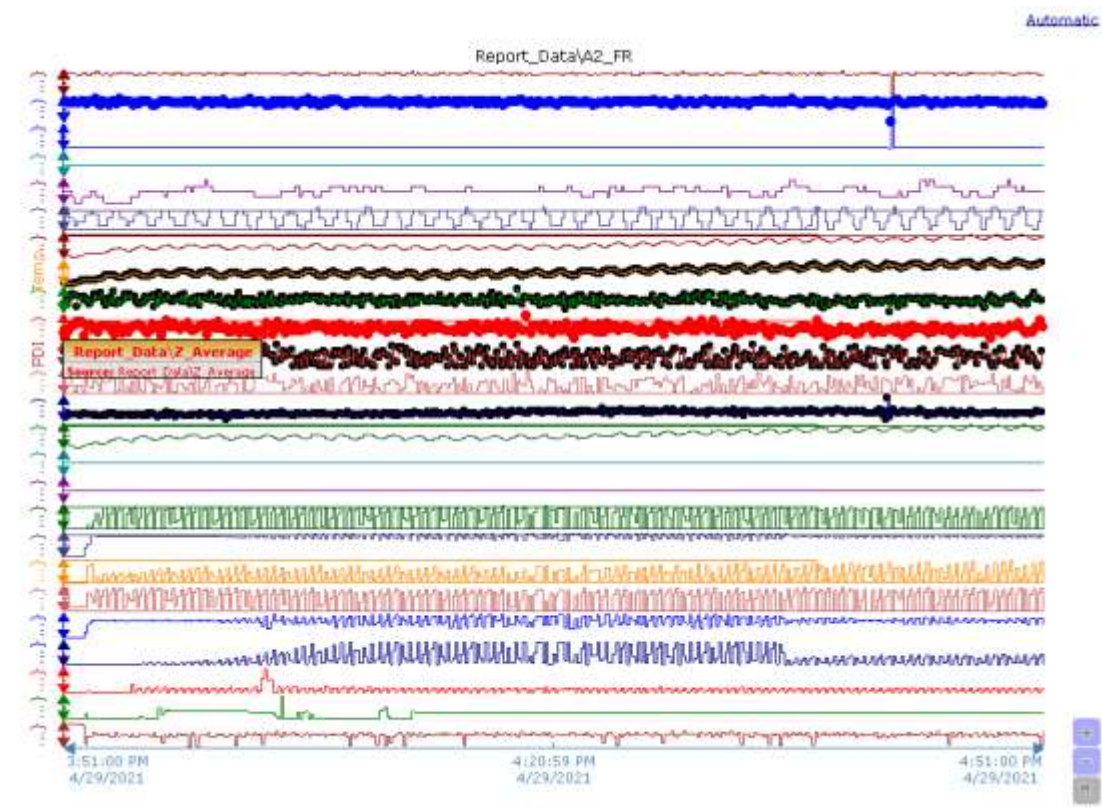
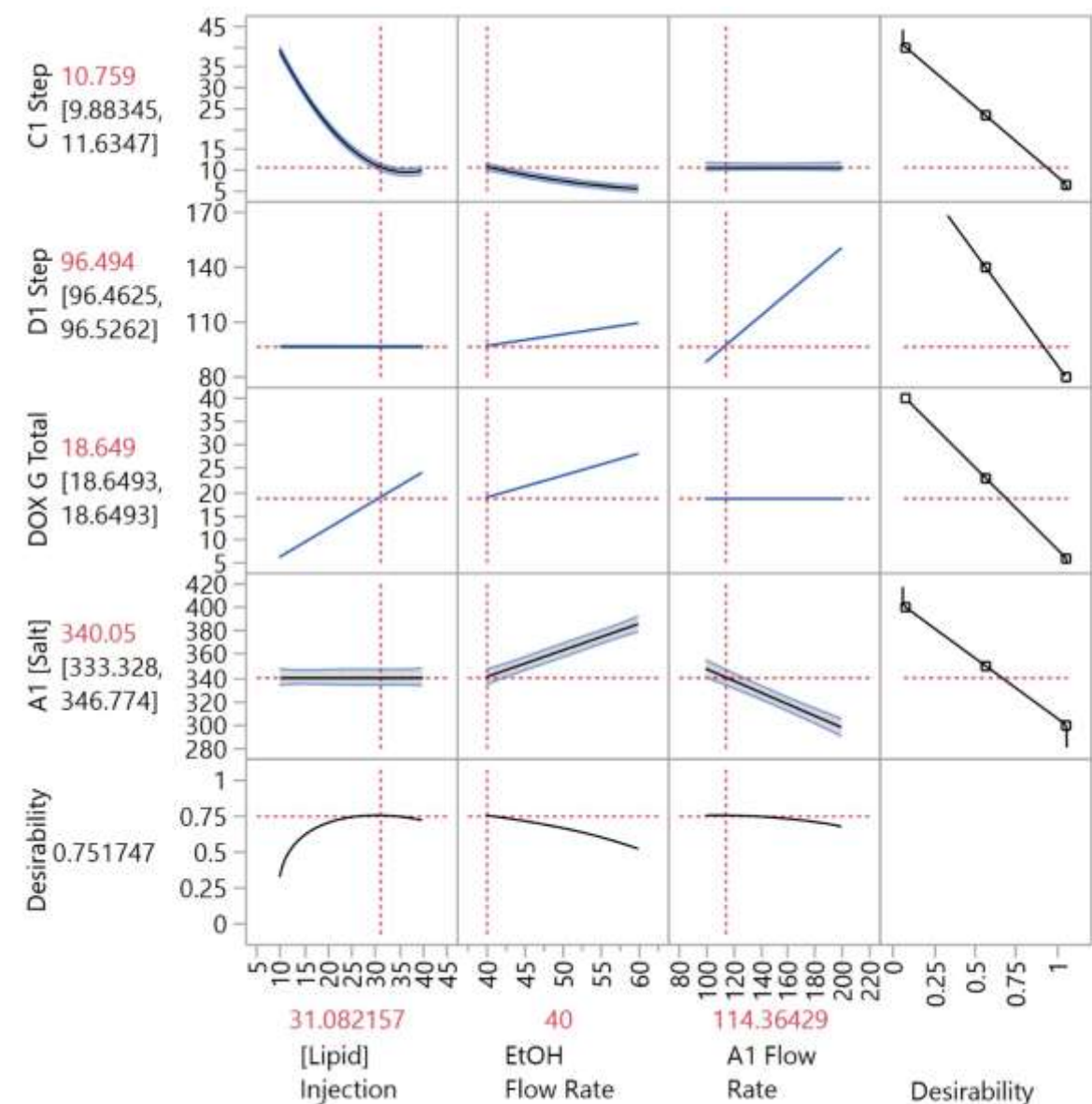


# Cryo-TEM of Liposomal Doxorubicin

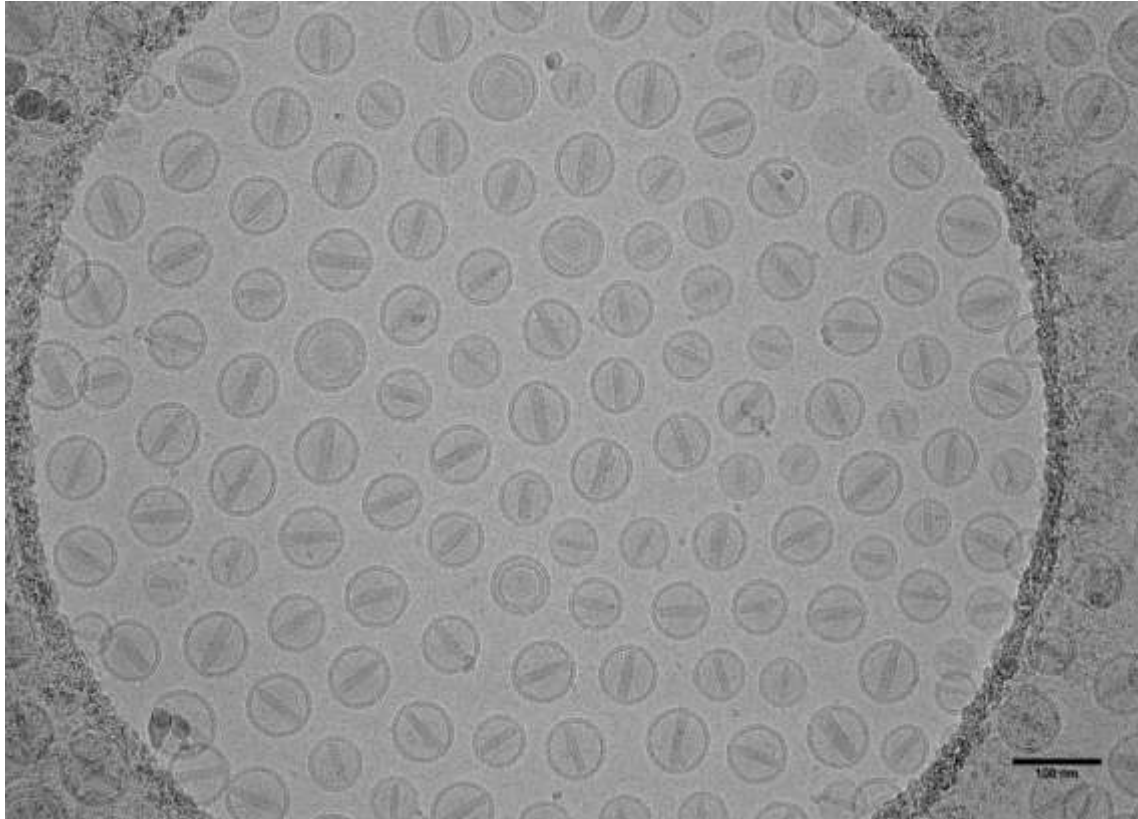


# Control Strategy: towards a working Design Space

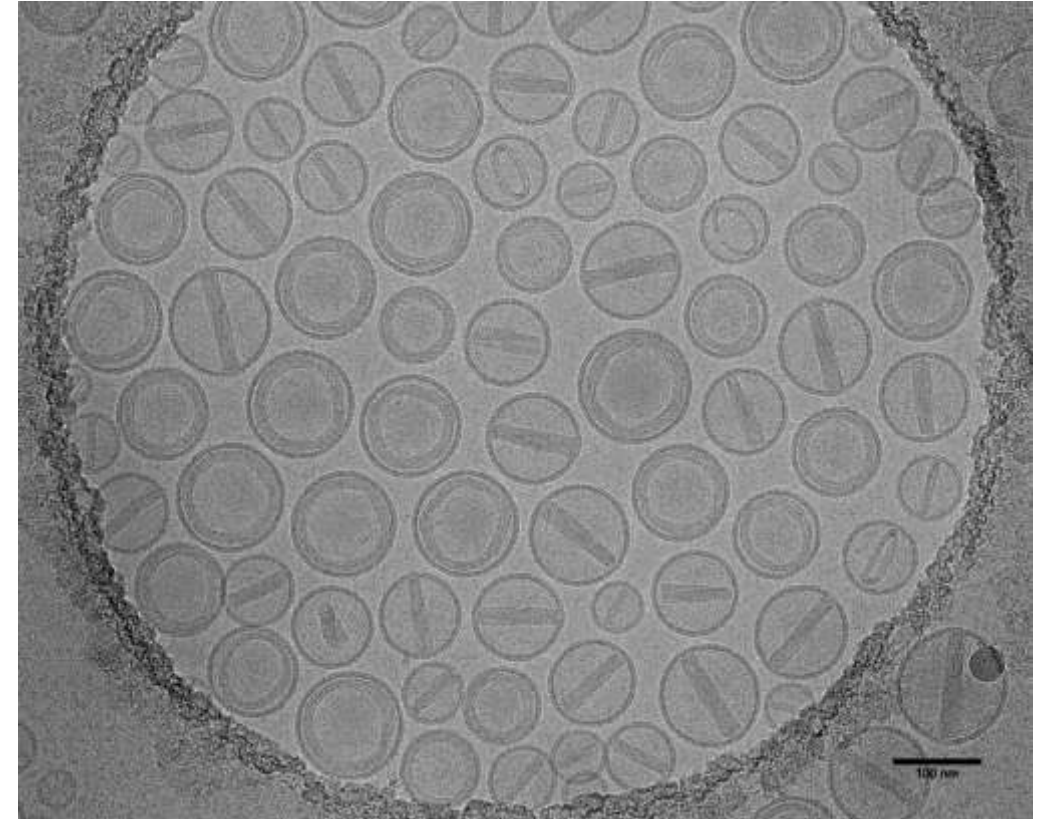
Use of Design of Experiments provides a framework to optimize conditions and run automation.



# Controlled API/ Crystal Morphology (Anti-cancer)



DLS: 86.68 d.nm



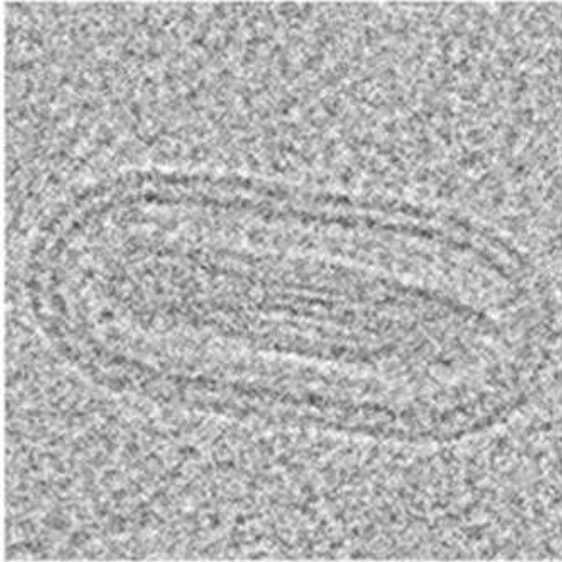
DLS: 123.33 d.nm

Linear crystal vs. spherical crystal  
For small-molecule anti-cancer therapies

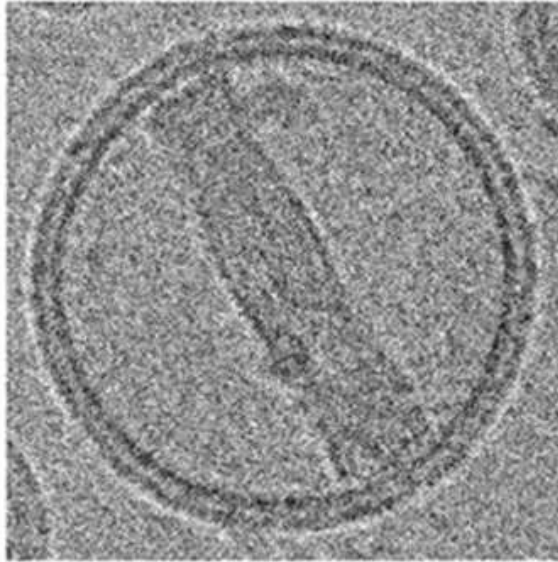


# Structures in Liposomal Doxorubicin

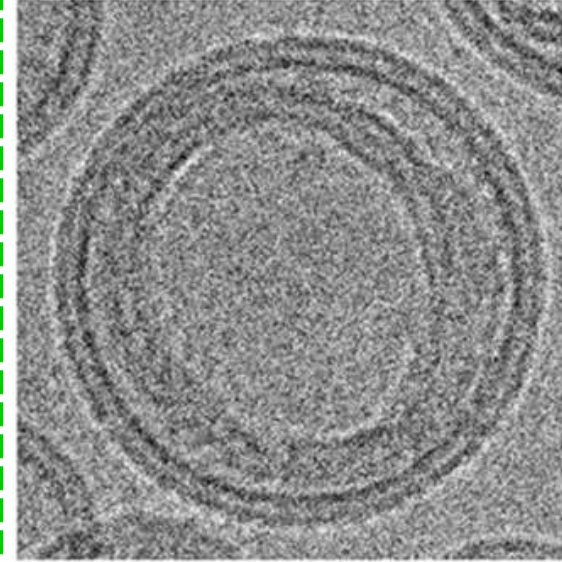
Majority of Particles



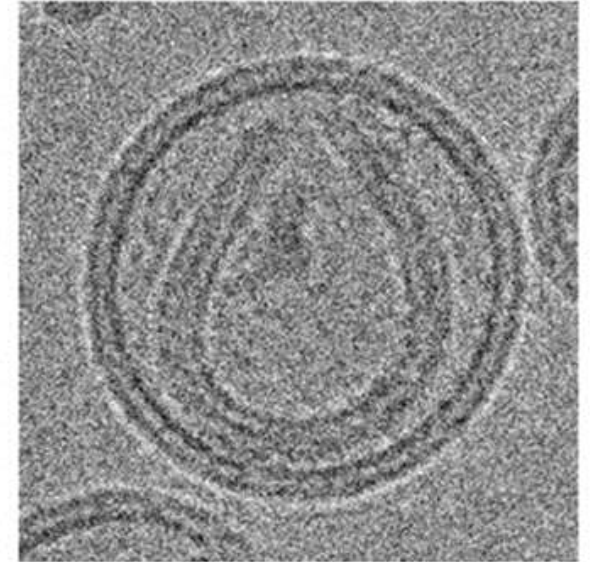
Elongated, Linear  
Nanocrystal



Spherical, Linear  
Nanocrystal



Spherical, Circular  
Nanocrystal

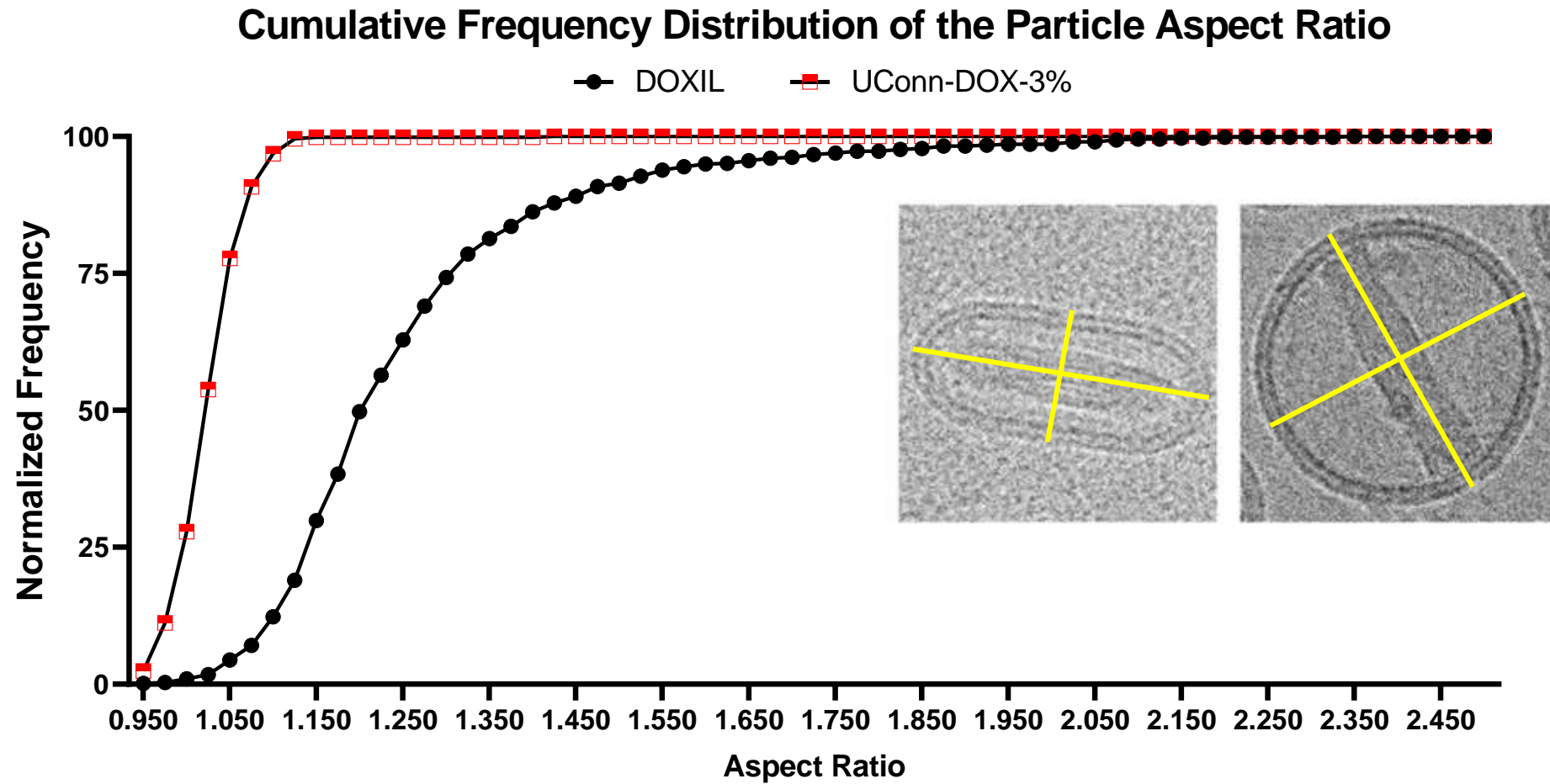


Spherical, U-Shaped  
Nanocrystal

Crystal shape impact on clinical adverse events? PK Data?

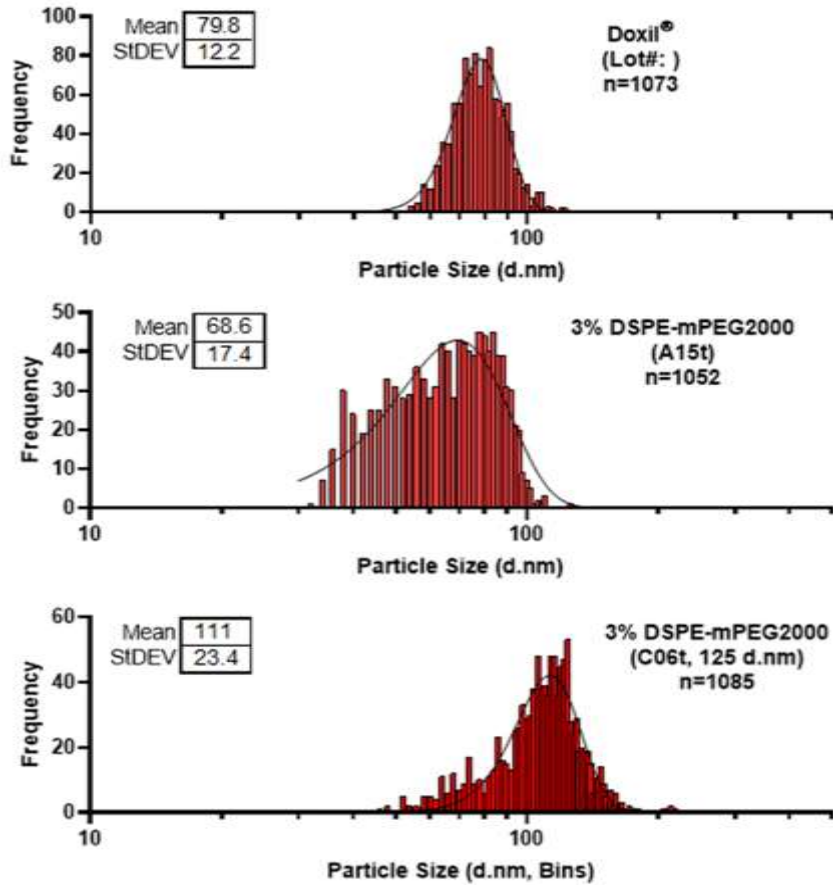


# Aspect Ratio of Salt Crystal

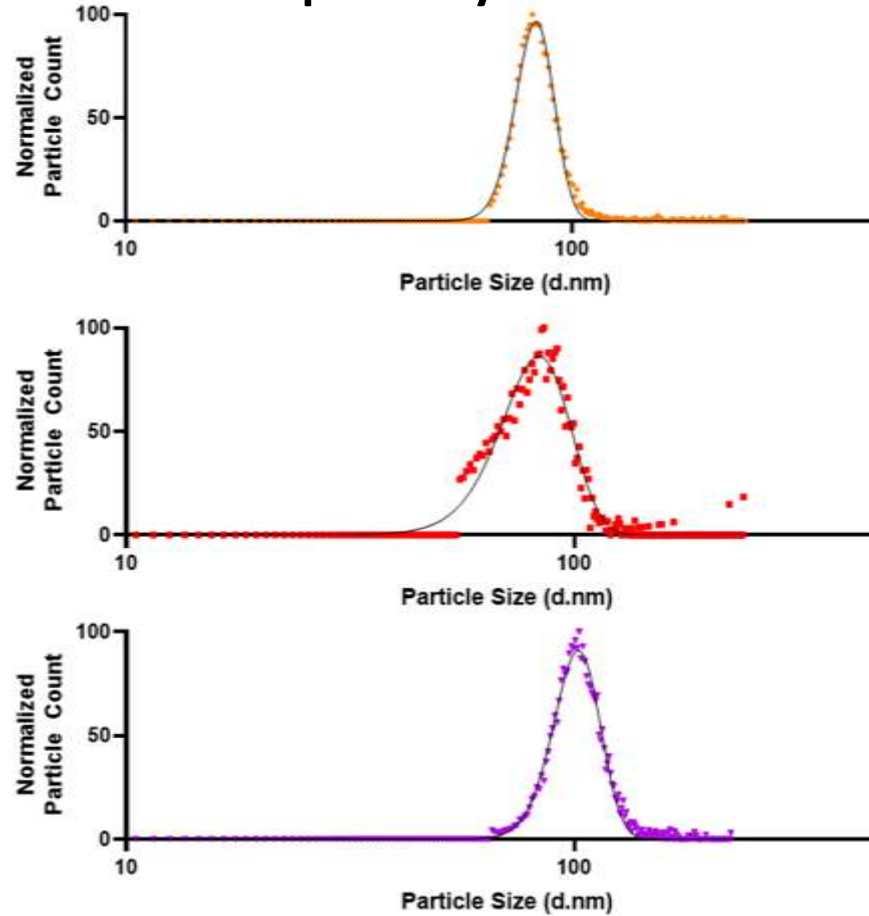


# Orthogonal Sizing Techniques

## Cryo-TEM (Number-Distribution)



## Microfluidic Resistive Pulse Sensing (Volume Distribution) Spectradyne nCS1



## DLS (Cumulant, Intensity-Based)

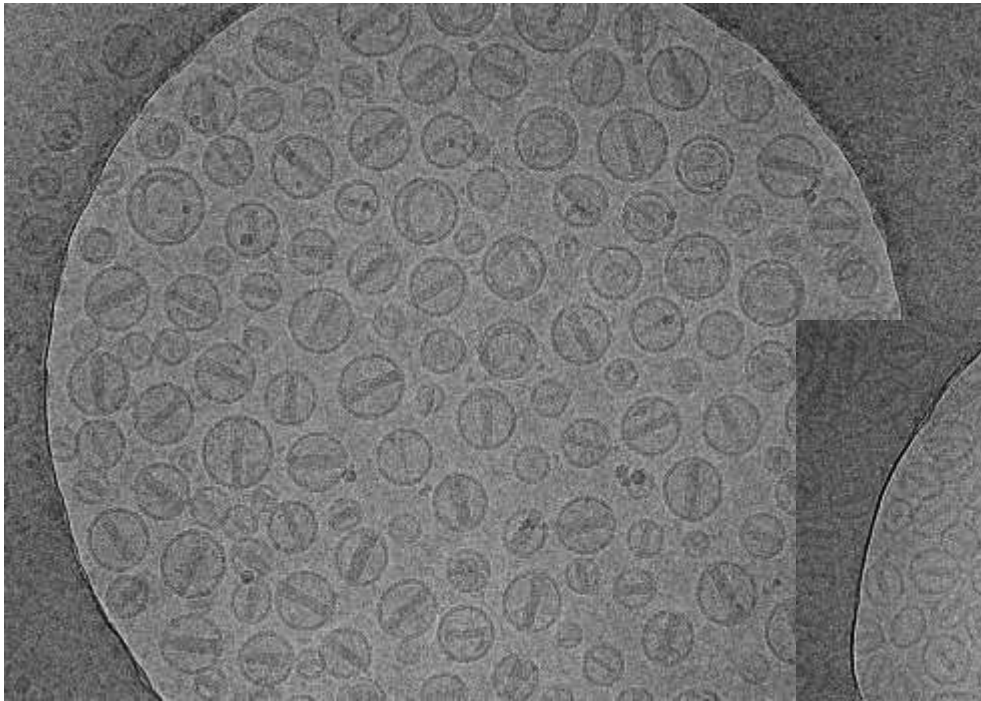
**Z-average** = 83.9 d.nm  
**PDI<sub>633</sub>** = 0.029

**Z-average** = 84.6 d.nm  
**PDI<sub>633</sub>** = 0.064

**Z-average** = 124.9 d.nm  
**PDI<sub>633</sub>** = 0.029

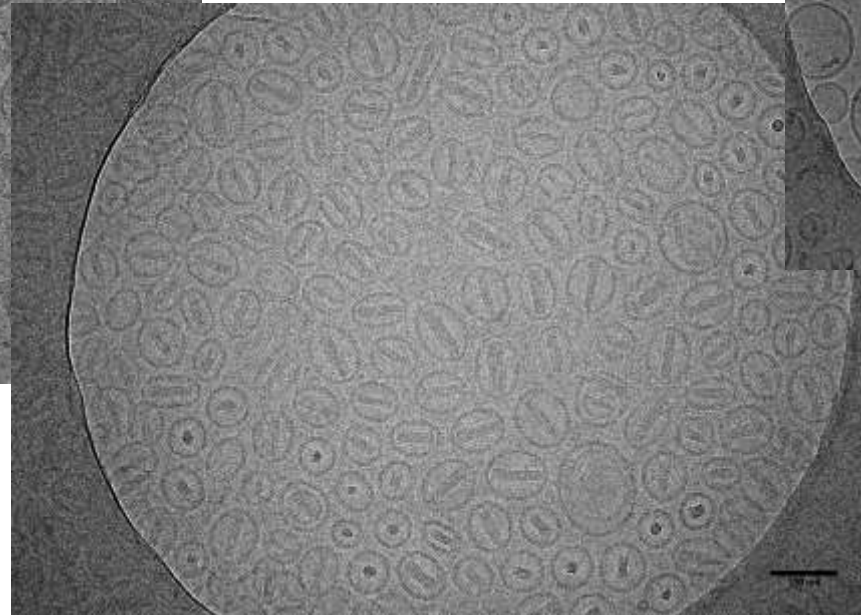
# Particle Size: Polydispersity and Wavelength

**PDI<sub>633</sub> = 0.064**  
**PDI<sub>1300</sub> = 0.107**

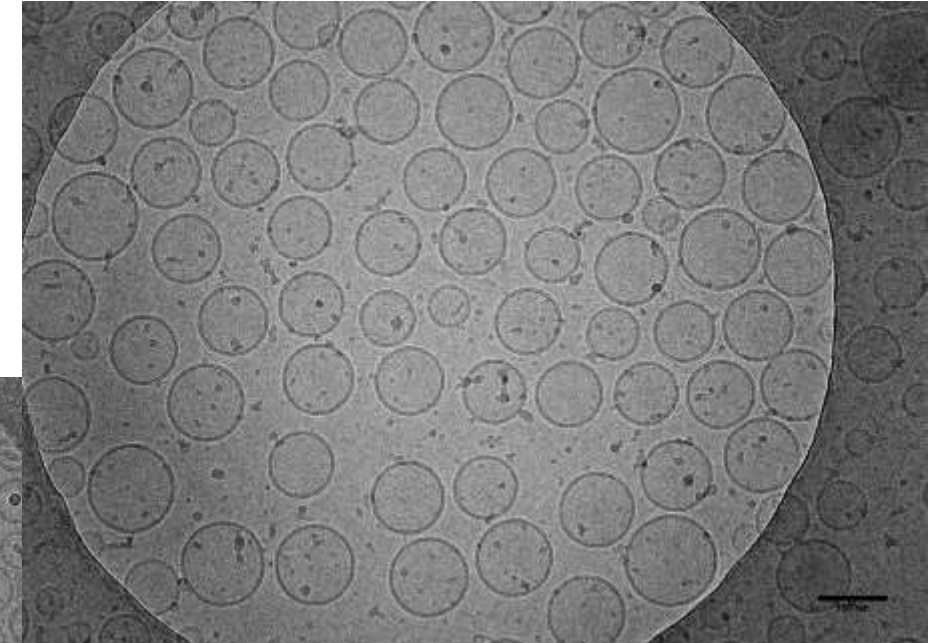


Size impacts on  
clinical adverse  
events?

**PDI<sub>633</sub> = 0.029**  
**PDI<sub>1300</sub> = 0.147**



**PDI<sub>633</sub> = 0.039**  
**PDI<sub>1300</sub> = 0.008**



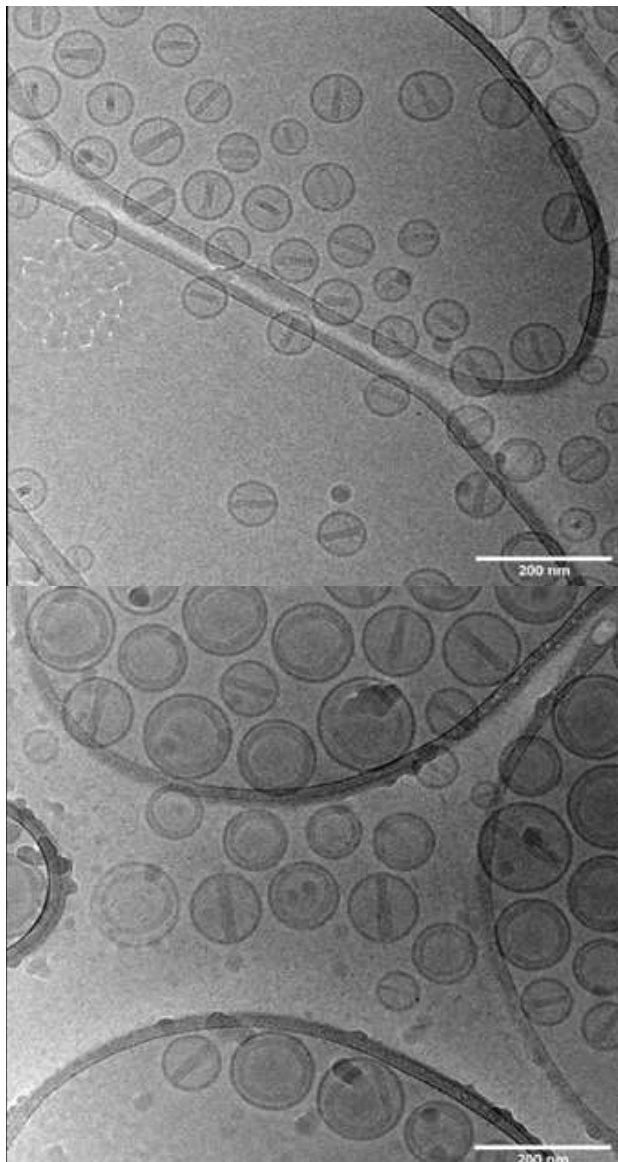
Should a Cumulant  
PDI  $\geq 0.10$  be acceptable  
for nanoparticles?

# Liposomal Doxorubicin In-Vivo Study

In collaboration with: Nanotechnology  
Characterization Laboratory, Frederick National  
Laboratory for Cancer Research



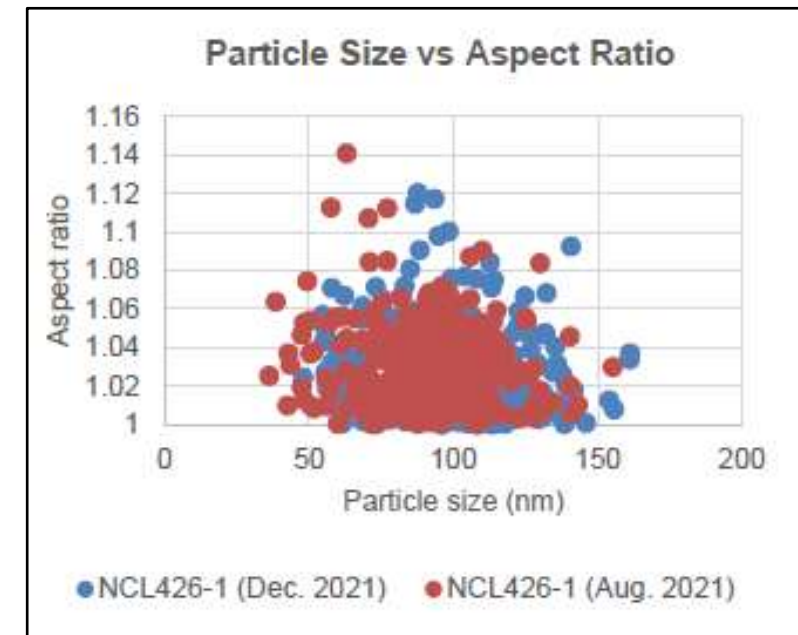
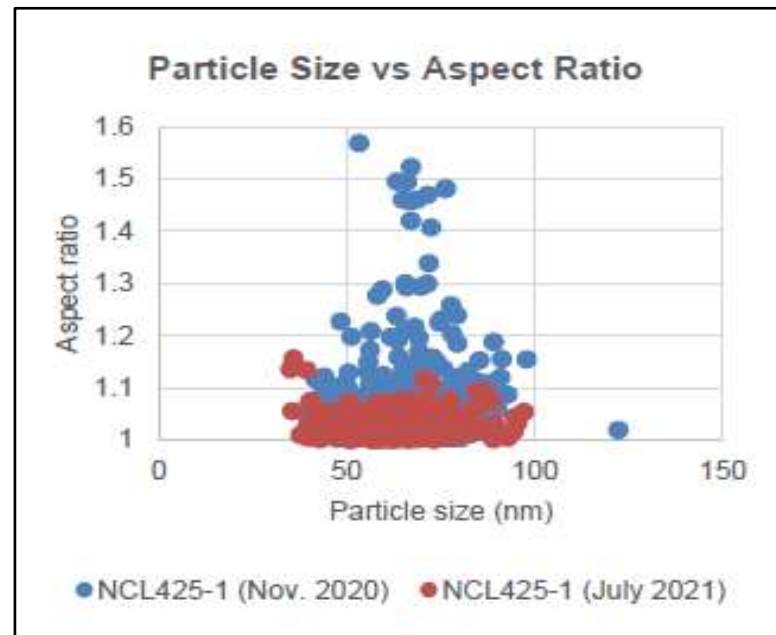
# Particle Characterization



NCL425

NCL426

Sample	Avg Diameter (nm)	Aspect Ratio	Linear Crystals (%)	Circular Crystals (%)
NCL425	$66.5 \pm 11.8$	$1.09 \pm 0.10$	86	10
NCL426	$102.6 \pm 21.1$	$1.02 \pm 0.02$	42	52



# Particle Size Characterization

## Hydrodynamic Size/Size Distribution using Dynamic Light Scattering

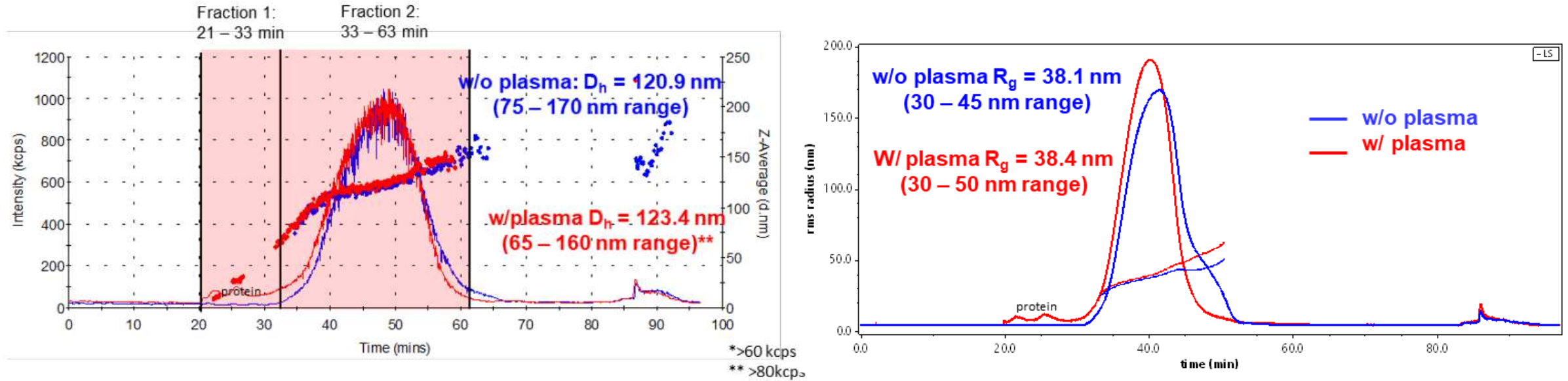
Sample	Dilution		Z-Avg (d.nm)	Int. Peak (nm)	PDI
NCL425	10 mM NaCl	100x	87.6 ± 0.6	91.3 ± 0.6	0.016 ± 0.011
		1000x	87.5 ± 0.05	91.2 ± 0.5	0.014 ± 0.006
	PBS	100x	87.3 ± 0.5	90.9 ± 0.5	0.013 ± 0.009
		1000x	87.5 ± 0.4	91.2 ± 0.5	0.018 ± 0.012
NCL426	10 mM NaCl	100x	119.5 ± 0.7	124.4 ± 0.5	0.016 ± 0.011
		1000x	119.7 ± 0.7	124.8 ± 0.7	0.015 ± 0.011
	PBS	100x	117.7 ± 0.8	123.9 ± 0.8	0.031 ± 0.017
		1000x	118.0 ± 0.6	124.7 ± 1.1	0.039 ± 0.008

Z-Avg: intensity-weighted average. PDI: polydispersity index. Int. Peak: intensity-weighted average over the primary peak.

- NCL425 exhibited an average size of approximately 90 nm, while NCL426 had a larger size of approximately 125 nm
- The size of a well-known manufactured drug product, previously measured at approximately 90 nm, aligns with NCL425

# Formulation Stability: NCL-426

## AF4 separation with in-line MALS and DLS



$\rho = R_g/D_h$ , where  $\rho$  = shape factor

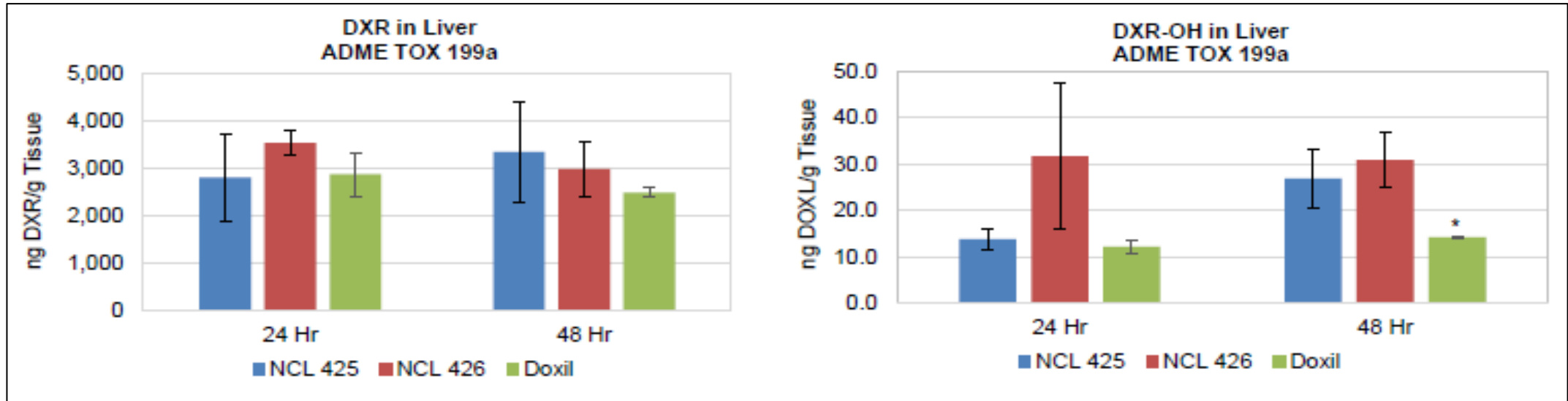
$$\rho_{\text{NCL426}} = 1.09$$

$$\rho_{\text{plasma}} = 1.11$$

$$\rho_{\text{Uniform sphere}} = 0.78$$

Minimal shift in  $\rho$ , suggesting minimal protein binding, consistent with manufactured drug product

# Toxicity Evaluation

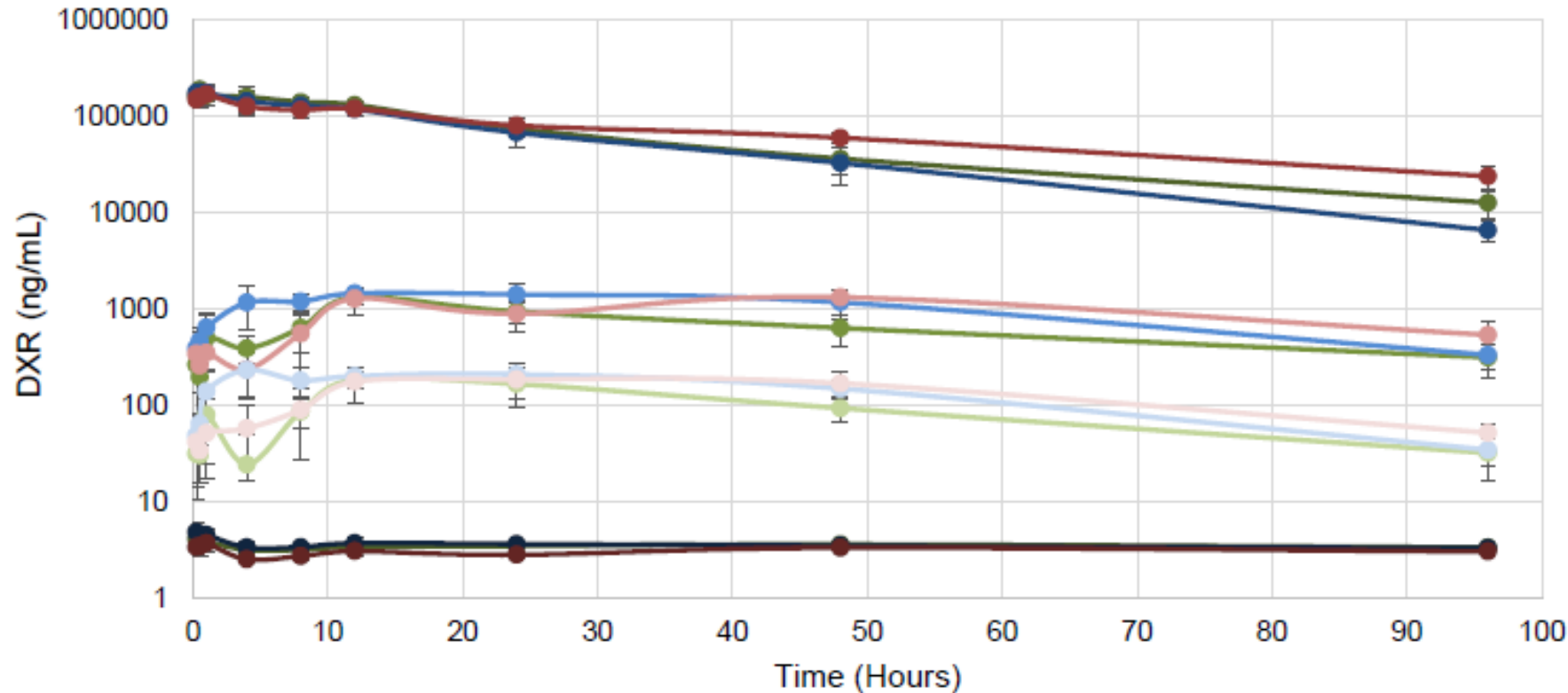


- Doxorubicin accumulation (left), and doxorubicinol accumulation (right) in the liver (Mean  $\pm$  SD, N=3).  
\*NCL426 vs. Mfd. Drug,  $p < 0.05$ , ANOVA with Tukey's post-hoc comparisons
- Doxorubicin accumulation in the heart, liver, and ear tissue was comparable among NCL425, NCL426, and manufactured drug product.
- Statistically significant differences were found in doxorubicinol concentrations in the heart and liver, indicating slight variations in tissue concentration profiles.



# Bioequivalence: In Vivo Drug Release

Total, Unencapsulated, and Unbound DXR and Doxorubicinol



NCL425, NCL426, and manufactured drug product, total, unencapsulated and unbound drug profiles were similar in vivo.

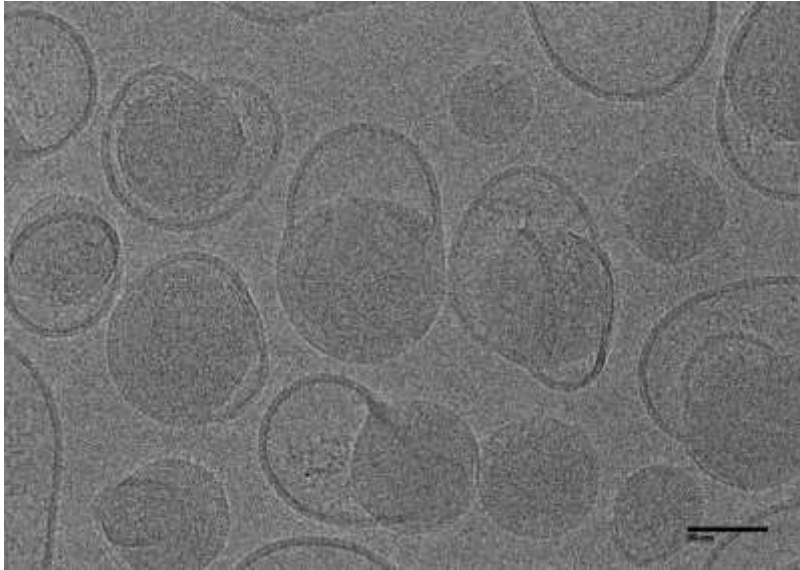
● NCL 425 Total	● NCL 426 Total	● Doxil Total
● NCL 425 Unencapsulated	● NCL 426 unencapsulated	● Doxil Unencapsulated
● NCL 425 Unbound	● NCL 426 Unbound	● Doxil Unbound
● NCL 425 Doxorubicinol	● NCL 426 Doxorubicinol	● Doxil Doxorubicinol

# mRNA-LNPs

## Structural Properties

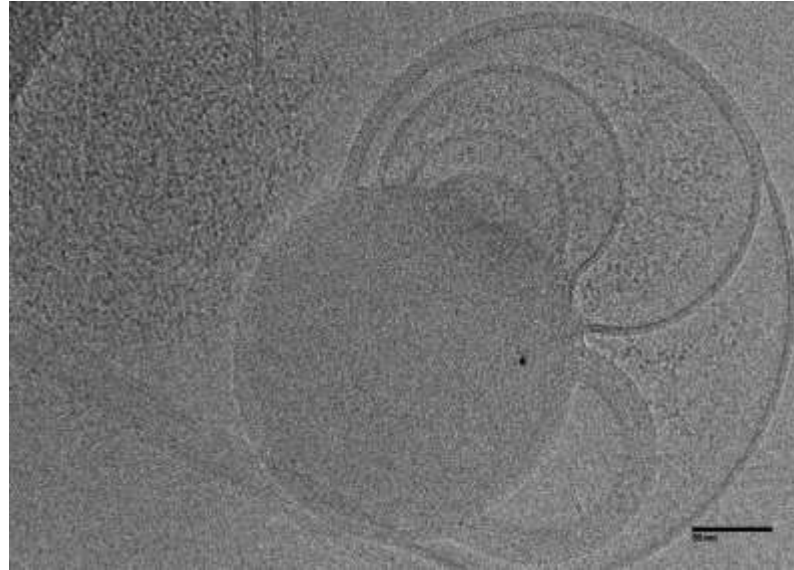
# mRNA-LNPs and formulation stability

**Low Phase Transition Lipid ,  
1.7% EtOH**



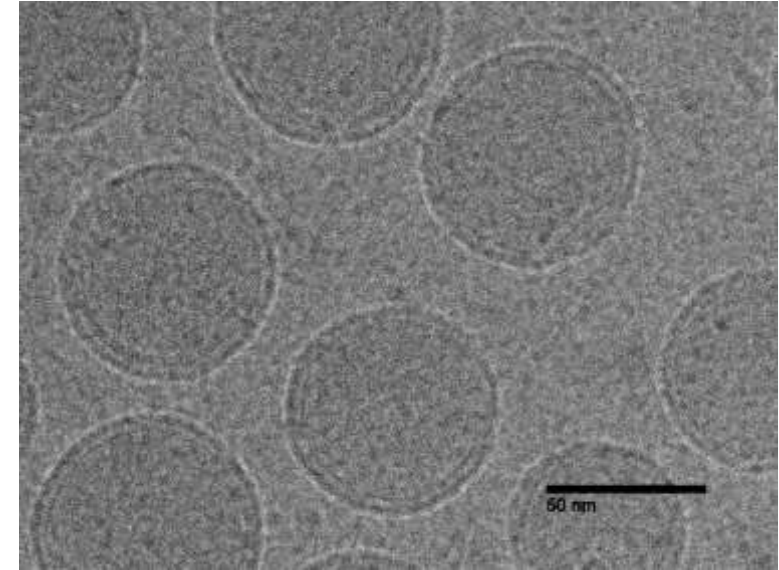
**Bleb-like structure  
Possible mRNA separation into  
multiple compartments**

**Low Phase Transition Lipid ,  
5.0% EtOH**



**Bleb-like structure, swelling,  
lamella extended  
Possible mRNA separation into  
multiple compartments**

**Higher phase transition lipids,  
ethanol <0.5%**

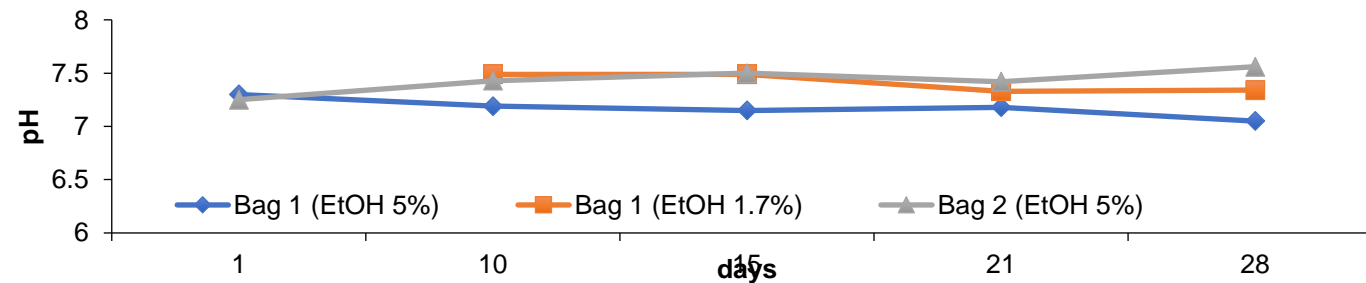
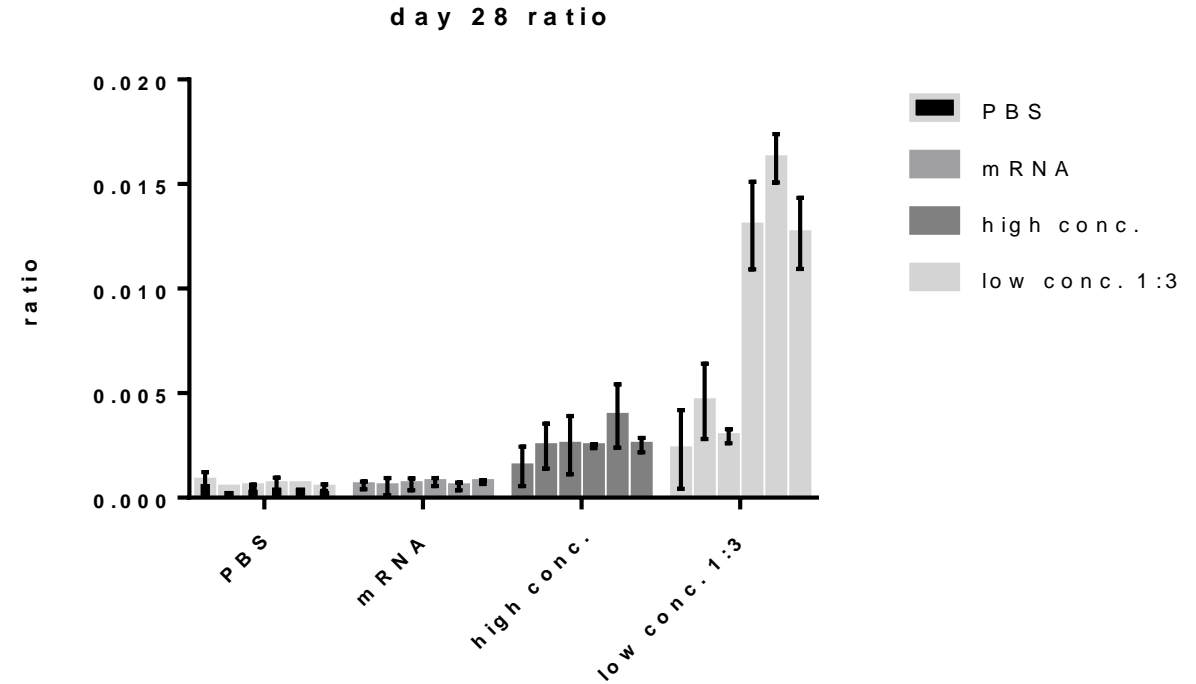
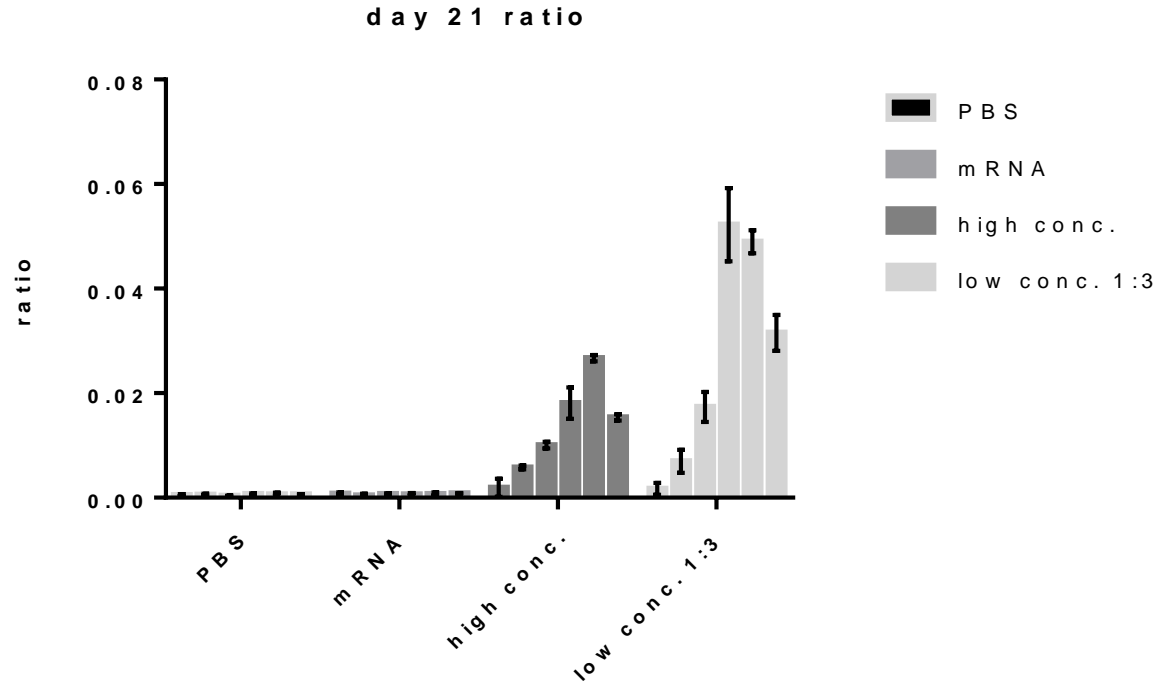


**Solid-core and no separation,  
more stable structure**

# mRNA LNP Transfection, 28 days

Particles: SSOP-POPE-Cholesterol-GM-PEG2k

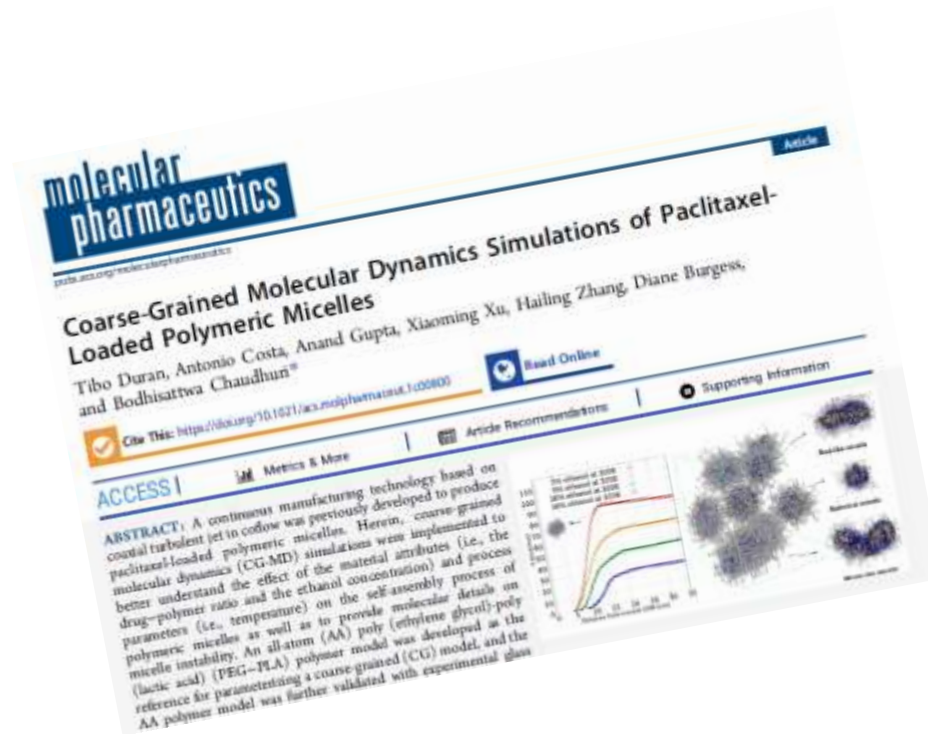
Cell line: K562, Chronic Myelogenous Leukemia Cells



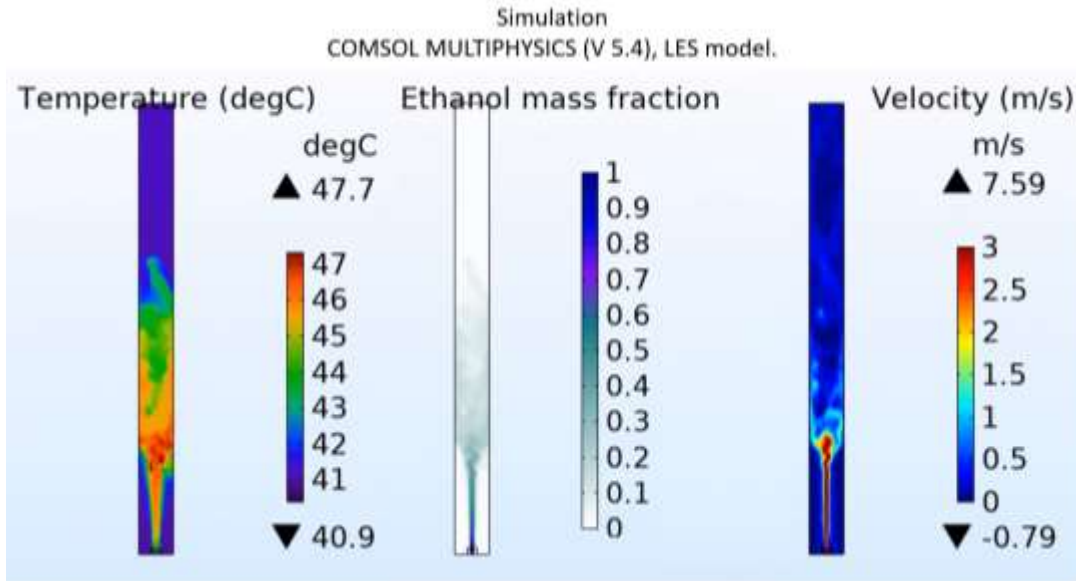


# Computational Fluid Dynamics and Molecular Dynamics (CG-MD)

*Work performed by Dr. Bodhi Chaudhuri's Lab*



# CFD Jet-flow Studies



Simulations provide additional information that is not measurable or difficult to measure.

## Eddy Current Analysis (Below)

Delta criterion defines vortices as regions in the value of delta is greater than 0.

$$\Delta = \frac{Q^3}{27} + \frac{R^2}{4} > 0$$

$\Omega_{ij}$ : rotation rate tensor

$S_{ij}$ : strain rate tensor

Experimental  
Track jet flow with blue dye in ethanol phase



100 mL/min  
33.6 °C



100 mL/min  
58 °C

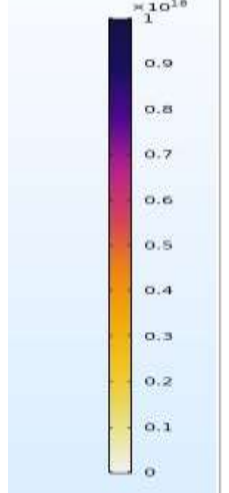


150 mL/min  
59.8 °C

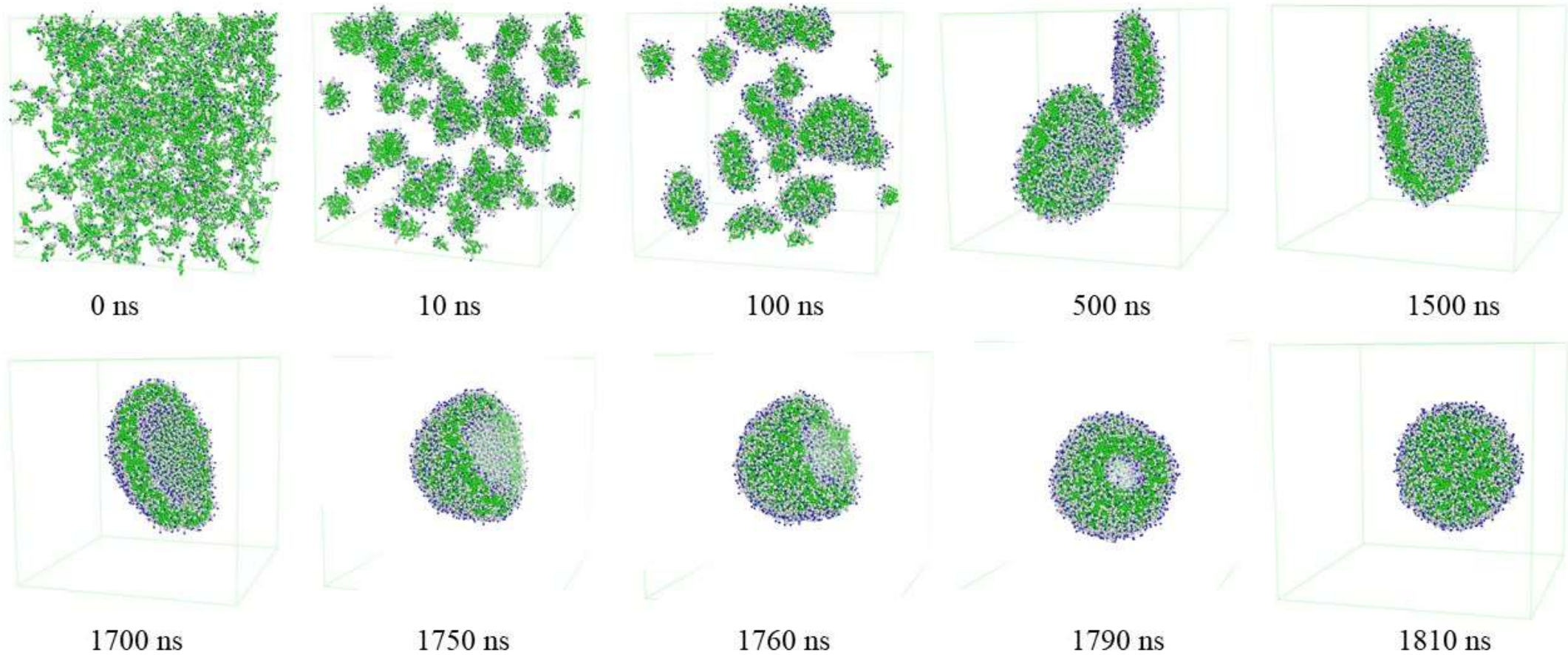


200 mL/min  
59.8 °C

Delta criterion  
 $\Delta$

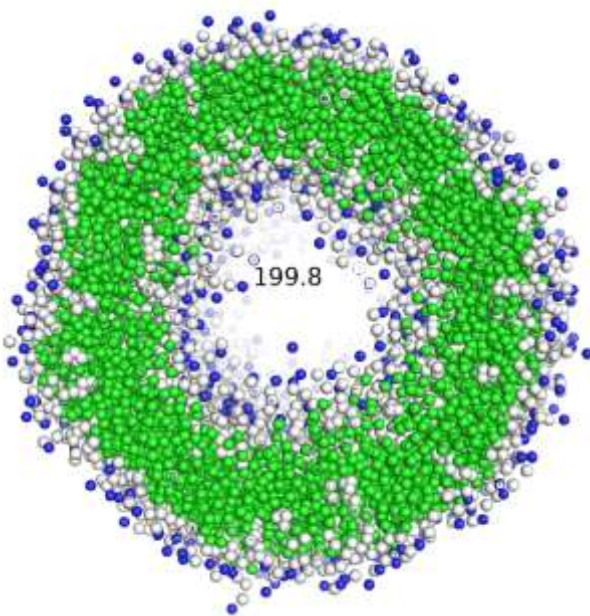


# MD Liposome Formation Studies

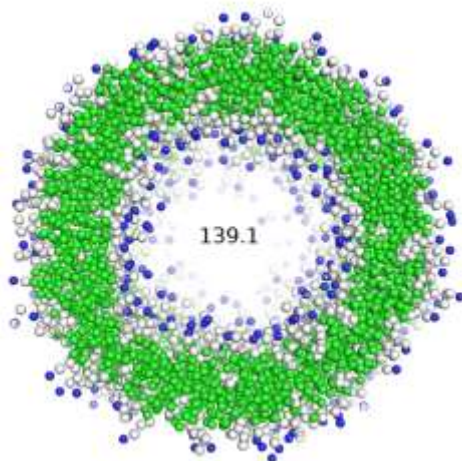


Simulation temperature: 333 K

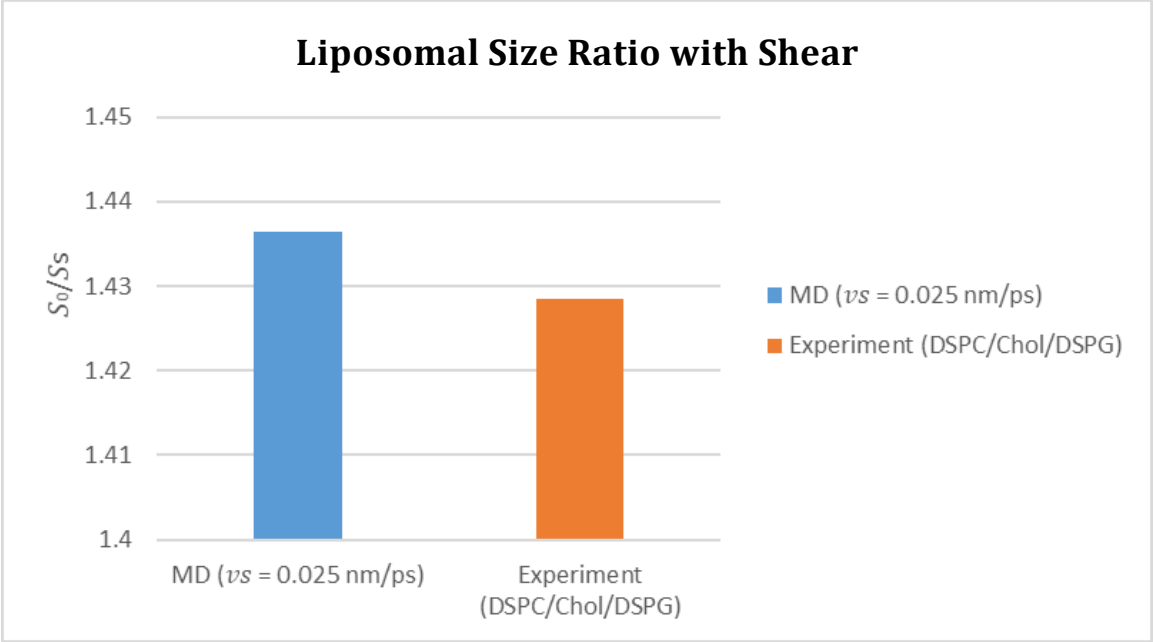
# Shear Simulations (Case $v_s = 0.025$ nm/ps)



Before shear (19.98 nm)



After shear (13.91 nm)



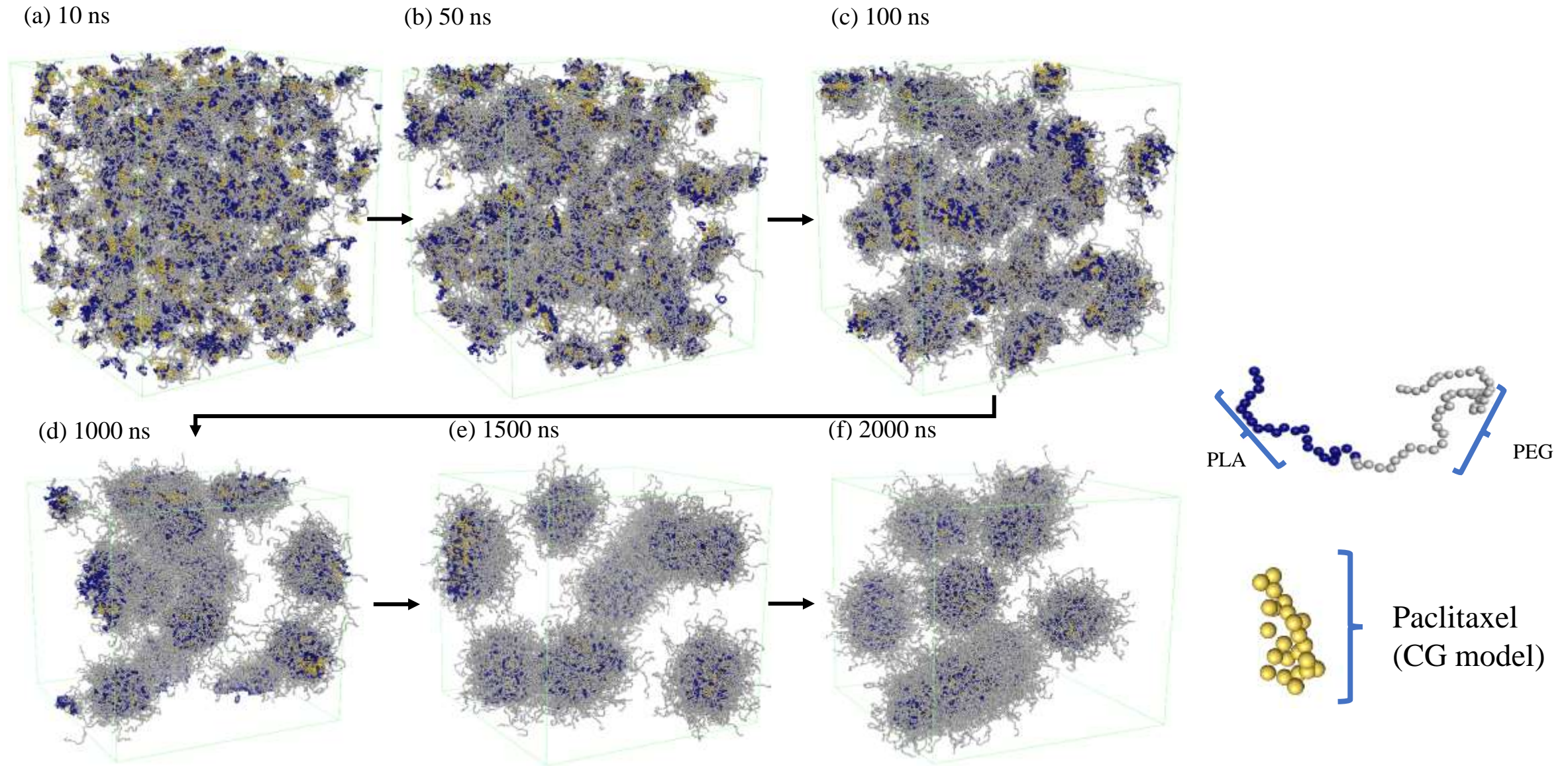
Liposome size ratio	Molecular Dynamics simulation results	Experiment results
$S_0/S_s$	1.436	1.428

- Higher shear rates resulted in smaller liposome formation
- MD issue is actual particle size due to constraints

$S_0$  = liposome size before shear  
 $S_s$  = liposome size after shear

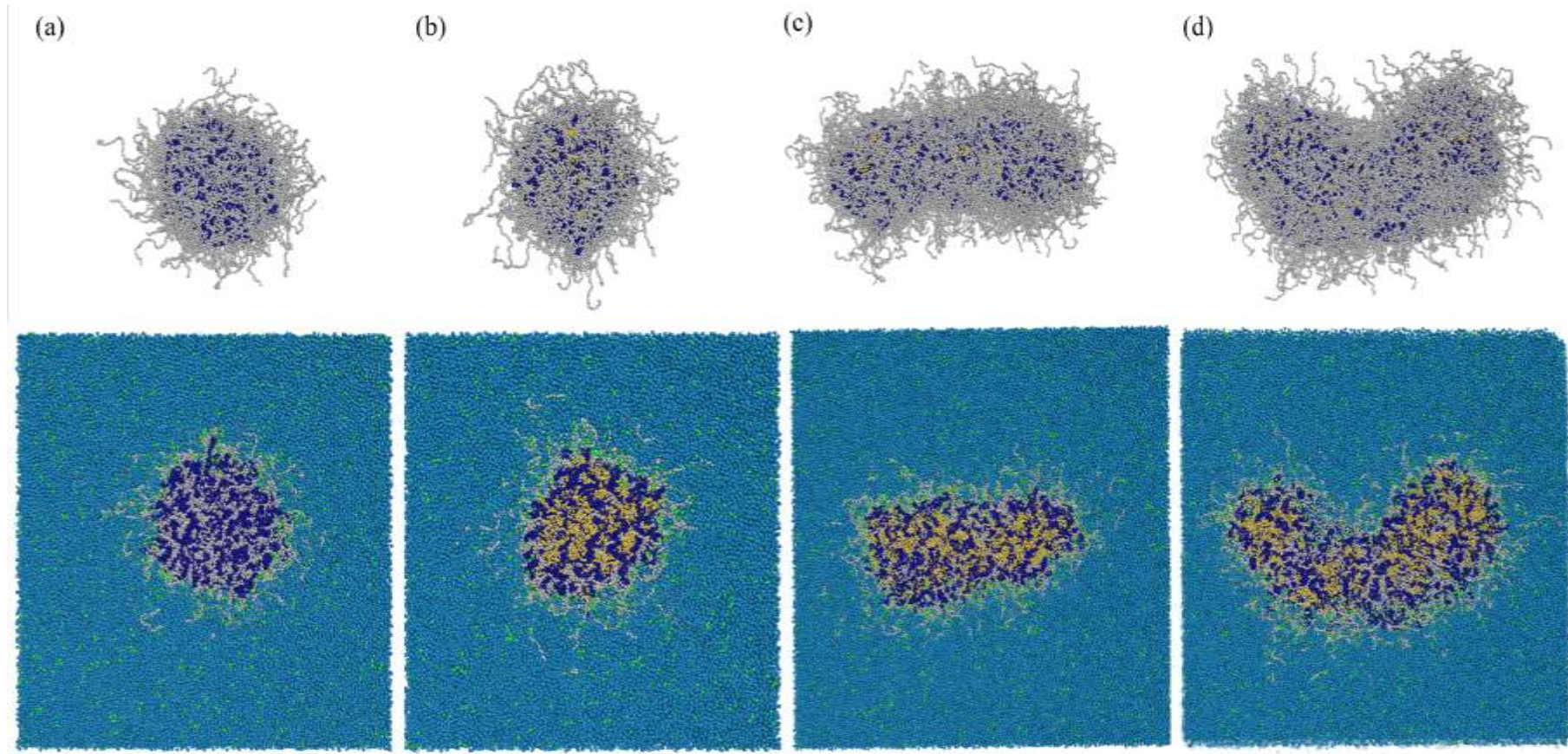


# Paclitaxel PEG-PLA Micelle Formation





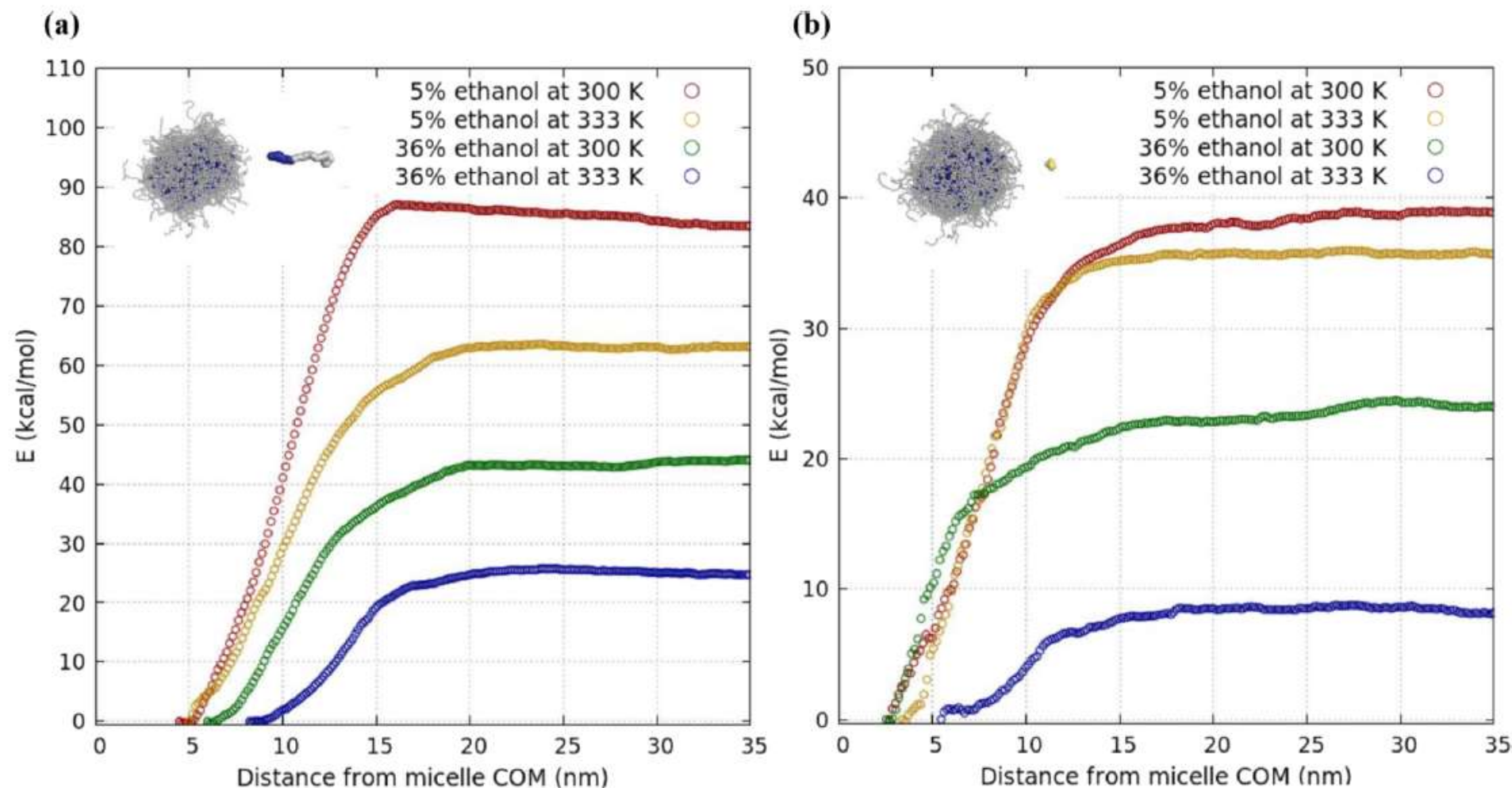
# Polymeric Micelle Structures



(a) blank spherical structures, (b) drug-loaded spherical structures, (c) drug-loaded rod-like structure, (d) drug-loaded worm-like structure

● PLA beads, ● PEG beads, ● PTX beads, ● water beads, and ● ethanol beads

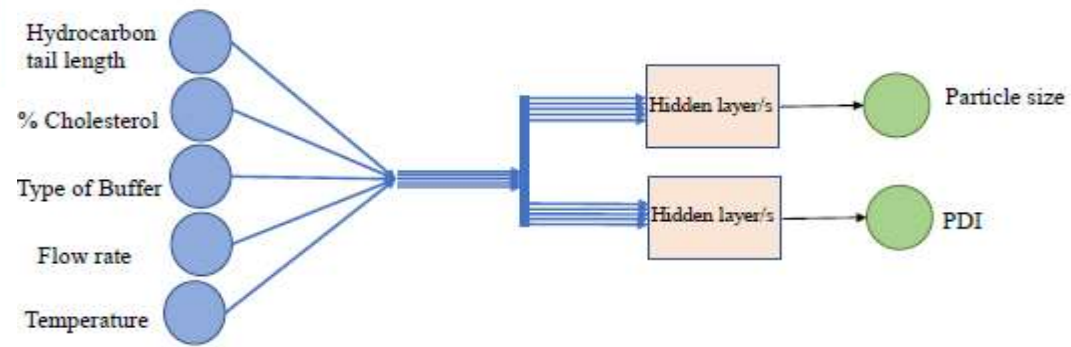
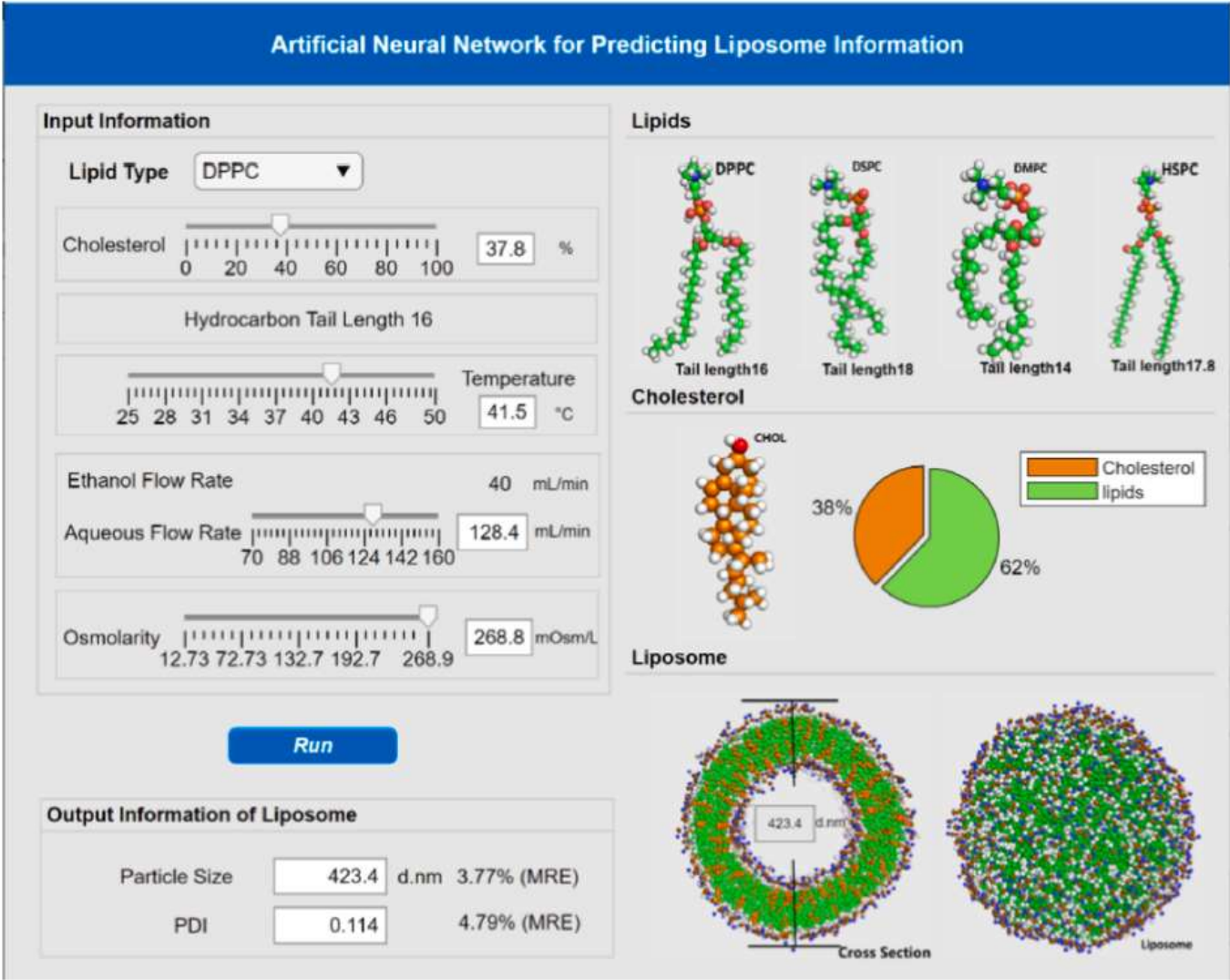
# Free Energy of Polymeric Micelle Dissociation



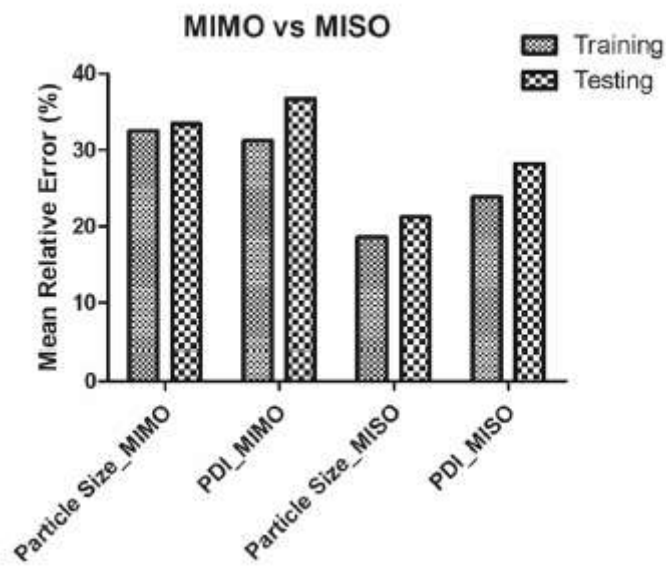
Potentials of the mean force calculated along the reaction coordinates for (a) pulling single PEG-PLA and (b) pulling single paclitaxel molecules away from the COM of the rest of the aggregate.



# Artificial neural networks for continuous manufacturing



Compared multi-input multi-output model (MIMO) and multi-input single-output model (MISO) models. MIMO vs MISO for predicting liposome particle size and PDI.





# Thank you!

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- 75F40120C00201: Continuous Processing of Liposomal Nanoparticles as Reference Materials for Drug Product (2020-2022)
- U01FD006975: Continuous Manufacturing of Nanoparticles: Establishing Real-Time-Release Testing Methods for a GMP-Ready System and Evaluation of Liposomal Morphological Changes in Real-Time (2020-2022)